



Review

Bioactive compounds and their libraries: An insight into prospective phytotherapeutics approach for oral mucocutaneous cancers

Henry A. Adeola^{a,b,1}, Afsareen Bano^{c,1}, Ravina Vats^c, Amit Vashishtha^d, Deepika Verma^e, Deepak Kaushik^f, Vineet Mittal^f, Md. Habibur Rahman^g, Agnieszka Najda^h, Ghadeer M. Albadraniⁱ, Amany A. Sayed^j, Sameh M. Farouk^k, Emad H.M. Hassanein^l, Muhammad Furqan Akhtar^m, Ammara Saleemⁿ, Mohamed M. Abdel-Daim^o, Rashmi Bhardwaj^{c,*}

^a Department of Oral and Maxillofacial Pathology, Faculty of Dentistry, University of the Western Cape and Tygerberg Hospital, Cape Town, South Africa

^b Division of Dermatology, Department of Medicine, Faculty of Health Sciences and Groote Schuur Hospital, University of Cape Town, Cape Town, South Africa

^c Centre for Medical Biotechnology, Maharshi Dayanand University, Rohtak, Haryana, India

^d Department Of Botany, Sri Venkateswara college, University of Delhi, India

^e Department Of Biochemistry, AllMS, Delhi, India

^f Department of Pharmaceutical sciences, Maharshi Dayanand University Rohtak, 124001, India

^g Department of Pharmacy, Southeast University, Banani, Dhaka 1213, Bangladesh

^h Department of Vegetable Crops and Medicinal Plants University of Life Sciences in Lublin 50A Doświadczalna Street, 20-280 Lublin, Poland

ⁱ Department of Biology, College of Science, Princess Nourah bint Abdulrahman University, Riyadh 11474, Saudi Arabia

^j Zoology Department, Faculty of Science, Cairo University, Giza 12613, Egypt

^k Cytology and Histology Department, Faculty of Veterinary Medicine, Suez Canal University, 41522 Ismailia, Egypt

^l Department of Pharmacology and Toxicology, Faculty of Pharmacy, Al-Azhar University, Assiut, Egypt

^m Riphah Institute of Pharmaceutical Sciences, Riphah International University, Lahore Campus, Pakistan

ⁿ Department of Pharmacology, Faculty of Pharmaceutical Sciences, Government College University Faisalabad, Faisalabad, Pakistan

^o Pharmacology Department, Faculty of Veterinary Medicine, Suez Canal University, Ismailia 41522, Egypt



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ABSTRACT

Oral mucocutaneous cancers (OMCs) are cancers that affect both the oral mucosa and perioral cutaneous structures. Common OMCs are squamous cell carcinoma (SCC), basal cell carcinoma (BCC) and malignant melanoma (MM). Anatomical similarities and conventions which categorizes these lesions blur the magnitude of OMCs in diverse populations. The burden of OMC is high in the sub-Saharan Africa and Indian subcontinents, and the cost of management is prohibitive in the resource-limited, developing world. Hence, there is a pressing demand for the use of cost-effective in silico approaches to identify diagnostic tools and treatment targets for diseases with high burdens in these regions. Due to their ubiquitousness and accessibility, the use of therapeutic efficacy of plant bioactive compounds in the management of OMC is both appropriate and plausible.

Abbreviations: OMC, Oral mucocutaneous cancer; SCC, Squamous cell carcinoma; BCC, Basal cell carcinoma; MM, Malignant melanoma; FDA, Food and Drug Administration; NCCIH, National Center for Complementary and Integrative Health; NCI, National Cancer Institute; HPLC, High-performance liquid chromatography; GC-MS, Gas Chromatography Mass Spectrometry; TLC, Thin layer chromatography; MS, Mass spectrometry; NMR, Nuclear magnetic resonance; COX-2, Cyclooxygenase-2; COPD, Chronic obstructive pulmonary disease; HT-1, Hepatorenal tyrosinemia-1; D2, Dopamine receptors; Hsp90, Heat shock protein 90; PI3K, Phosphoinositide 3-kinase; PKB, Protein kinase B; MAPK, Mitogen-activated protein kinase; NF-κB, Nuclear factor kappa B; IL-8, Interleukin 8; IL-6, Interleukin 6; TNFα, Tumor necrosis factor-alpha; AMPK, AMP-activated protein kinase; EGFR, Epidermal growth factor receptor.

* Corresponding author.

E-mail addresses: henry.adeola@uct.ac.za (H.A. Adeola), afsareen.rs.cmbt@mdurohtak.ac.in (A. Bano), ravina.rs.cmbt@mdurohtak.ac.in (R. Vats), vashishtha24@svc.ac.in (A. Vashishtha), drdeepikaverma@aaims.edu (D. Verma), deepkaushik.pharma@mdurohtak.ac.in (D. Kaushik), drvineet.pharma@mdurohtak.ac.in (V. Mittal), pharmacisthabib@gmail.com (Md.H. Rahman), agnieszka.najda@up.lublin.pl (A. Najda), gmalbadrani@pnu.edu.sa (G.M. Albadrani), amanyasayed@sci.cu.edu.eg (A.A. Sayed), dr_smf_hist@vet.suez.edu.eg (S.M. Farouk), emadhassanien@azhar.edu.eg (E.H.M. Hassanein), mfurqan.akhtar@riphah.edu.pk (M.F. Akhtar), ammarasaleem@gcu.edu.pk (A. Saleem), abdeldaim.m@vet.suez.edu.eg (M.M. Abdel-Daim), bhardwajrashmi.cmbt@mdurohtak.ac.in (R. Bhardwaj).

¹ These authors contributed equally to this work.

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Furthermore, screening known mechanistic disease targets with well annotated plant bioactive compound libraries is poised to improve the routine management of OMCs provided that the requisite access to database resources are available and accessible. Using natural products minimizes the side effects and morbidities associated with conventional therapies. The development of innovative treatments approaches would tremendously benefit the African and Indian populace and reduce the mortalities associated with OMCs in the developing world. Hence, we discuss herein, the potential benefits, opportunities and challenges of using bioactive compound libraries in the management of OMCs.

1. Introduction to bioactive compound libraries and product synthesis

The traditional use of in-vitro physical screening of large bioactive compound libraries for drug discovery is laborious and is being replaced by computer-aided virtual approaches, albeit the structural diversity and biological activities of natural product are inimitable [1]. Bioactive compound libraries are families of natural products and can be defined as a collection of pre-certified, functionally diverse structures, obtained from different natural sources, that have a wide range of applications [2]. Chemical reactions that are well-characterized are employed to create these highly valuable set of library compounds from easily available monomers, from which library compounds can be designed using a combinatorial approach [3]. However, high-quality screening of large bioactive compound libraries for RNA or protein in an unbiased way was a limitation of combinatorial chemistry; and the “rise and fall of combinatorial chemistry” has resulted in an increased use of in-silico virtual compound libraries [4]. Furthermore, the lack of specificity of the bioactive compound in therapeutic targeting of cellular/subcellular sites, fraught their routine uses [5].

These libraries include a vast array of products such as, natural products, pioneering compounds, approved compounds, and clinical compounds [6]. They can also be used for signal pathway research, drug discovery and drug repositioning [7]. Bioactive compounds are of immense importance in the management of various medical conditions/diseases, including cancer [8]. These natural compounds, which are largely isolated from microorganisms, plants and animals are bioactive and exert pharmacological effects in the treatment of human pathologies. Furthermore, these bioactive compounds can be used as nanocarriers for drug-delivery due to their superior biocompatibility, as compared to synthetic nanoparticles [8]. Virtual phenotypical screening of bioactive compound libraries is a cost-effective, seamless and plausible approach to the discovery of drug targets in human diseases. Although, these compound libraries present a treasure trove of targeted

therapies for various medical conditions, their use has been poorly explored in the field of dentistry and oral pathology, particularly in oral mucocutaneous cancers (OMCs). Hence, this review seeks to discuss the application of natural and virtual compound libraries in the management of common OMCs viz: basal cell carcinomas (BCC), malignant melanomas (MM) and squamous cell carcinomas (SCC). Currently, there are six major orthodox therapeutic interventions for mucocutaneous cancers viz; surgery, radiation therapy, chemotherapy, immunotherapy, targeted therapy, Bone marrow/stem cell transplant and hormone therapy. Despite progress made with these interventions, the use of natural bioactive compounds and compound libraries has a tremendous complementary benefit as an ancillary tool for OMC cancer management.

2. Physical natural product libraries and products

Several microbes, animals and plants with bioactivity can be candidates for the compound library development (Fig. 1). Various parts of plants such as stem, root, leaf, fruit and flower as well as microbial cultures are also vital sources for bioactive compound extraction using various approaches such as, molecule distillation, membrane separation, ultracentrifugation and enzyme-mediated extraction techniques. The products obtained are then safely stored in Natural Compound Libraries which are employed for drug discovery applications. These bioactive compound libraries have certain features in common as enlisted below:

- All necessary safety measures to ensure safety and bioactivity of natural extracts are established by various preclinical and clinical trials.
- All the important details of natural compound, for instance- chemical structure, physical properties, IC₅₀ value etc., are reported.
- Bioactive compounds or extracts provided by these libraries are used in research work concerned with drug discovery for various diseases such as neurological disorders, cancer, cardiac disorders etc.

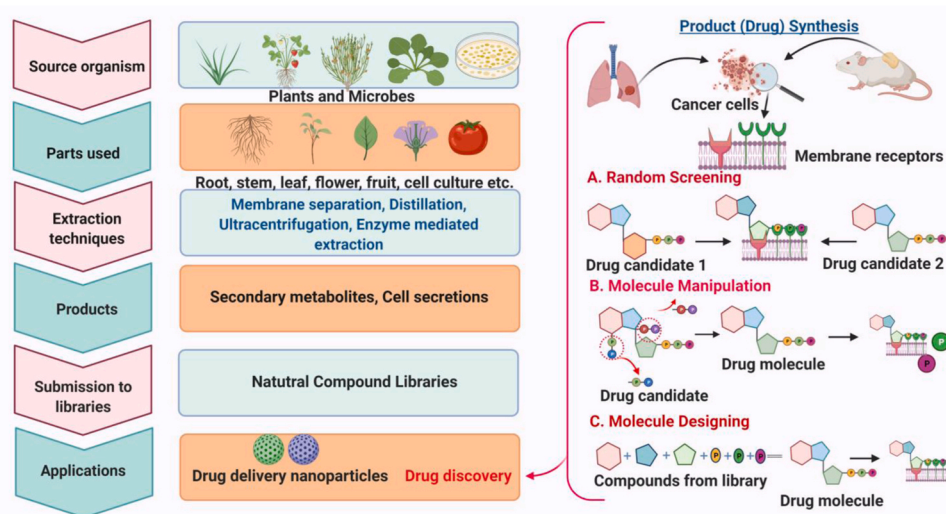


Fig. 1. Natural Compound Libraries and Product. Explanatory notes: Various plants and microbes with medicinal properties can be used as a source for the extraction of drug candidates. Root, stem, leaf, flower, fruits and microbial cell cultures are the starting materials that are subjected to different extraction techniques such as, membrane separation, molecule distillation, ultracentrifugation, enzyme mediated extraction techniques. Products obtained through these techniques are stored/kept safe in Natural Compound Libraries. These compounds from Natural Compound Libraries are used for various applications including drug discovery and synthesis. (Created with www.BioRender.com).

- Apart from therapeutic effects, these compounds are actively used in various sectors such as agrochemicals, nutraceuticals, cosmetics, antimicrobials and insecticides manufacturing.
- The phyto-compounds available in these natural product libraries are used in discovery of drugs with low toxicity.
- These compounds or extracts are provided with prior approval by FDA.
- To confirm the accessibility of diverse bioactive compounds, these libraries are *ad infinitum* rationalized.

A list of such libraries of natural products has been supplied by

National Center for Complementary and Integrative Health (NCCIH) which serves principle idea to provide relatively comprehensive information (see Table 1). These biocompound libraries include natural as well as synthetic extracts not only from plants but from fungus, insects and other organisms also.

The successful execution of a precedential pipeline for efficient natural products depends on organized compilation of natural raw extracts. The most prevalent and diverse library for screening of natural extracts was evolved in the 1980's and 1990's by the National Cancer Institute (NCI) [9]. An enormous number of specimens and cultures from diverse natural sources were processed by NCI and other cancer research

Table 1

List of various libraries providing natural product extracts.

	Name of the Library/ Service provider	Date of Establishment	Founder/ CEO/ Managing Director	Headquarters/ Office	Web-address	Materials/Information/Data Available in the Library
1.	Albany Molecular Research Inc.	1991	Thomas E. D'Ambra	Albany, New York, USA	https://www.amriglobal.com/	Natural products obtained from marine and terrestrial microbes and plants
2.	Biosortia Pharmaceuticals	2012	Ross O. Youngs	San Diego, California, United States	https://www.biosortia.com/	Highly defined natural products extracted unswervingly from in-situ source i.e., aquatic micro biome.
3.	Caithness Biotechnologies Ltd	2015	Clett Erridge	Leicester, United Kingdom	http://caithnessbiotechnologies.com/	Inimitable focus on specific plants with medicinal properties. Risk management through dietary epidemiology. Endeavor to exploit accessibility of natural compound libraries. Approximately 800 extracts are available in DMSO, microplate format.
4.	ChromaDex®	1999	Frank L. Jaksch	Los Angeles, California, United States	https://www.chromadex.com/	Extensive medley of high-quality Natural Products/Compounds along with references. Appropriate for the Industries associated in the midst of food and beverages, pharmaceutical and cosmetic souk.
5.	Cyano Biotech	2004	Dan Kramer	Berlin, Germany	http://www.cyano-biotech.com/	Provide natural compounds obtained from cyanobacteria.
6.	Developmental Therapeutics Program	1955	NCI, NIH	USA	https://dtp.cancer.gov/	One of the comprehensive compilations of natural products. Natural compounds are extracted from different microorganisms and plant sources present all over the world. It also includes Library of Medicinal Plant Extracts from China.
7.	Greenpharma	2000	Philippe Bernard	Orléans, France	https://www.greenpharma.com/company/	Miscellaneous purified natural compounds from numerous sources like bacteria, plants etc.
8.	INDOFINE Chemical Company Inc.	1981	Kotesarama Bezwada	New Jersey, USA	http://www.indofinechemical.com/	This company is concerned with providing herbal and dietary products.
9.	InterBioScreen	1997	Dr. Kartsev	Chernogola, Russia	https://www.ibscreen.com/	Synthetic and natural compounds along with their derivatives from natural sources including plant, fungus, insects, marine organisms etc.
10.	InterLink Biotechnologies	1991	Dr. Garcia-Lazcano	Princeton, New Jersey, USA	http://www.interlinkbiotech.com/	Microbial and plant extracts.
11.	Magellan BioScience	1997	John M. Cronan	Tampa, Florida, United States	http://www.magellanbioscience.com/	Extracts from invertebrates and plants.
12.	MicroSource Discovery Systems Inc.	1993	John Devlin	New Milford, CT, US	http://www.msdiscovery.com/	Collection includes alkaloids, flavanoids, sterols, diterpenes, benzophenones, and coumarins.
13.	Natural Products Discovery Institute	2011	Merck and Company	California, USA	https://www.scripps.edu/support-us/natural-products/	Microbial fermentations, plant sources, raw and partitioned extracts
14.	NatureBank	2015	James Tansey	Vancouver, Canada.	https://www.naturebank.com/	Marine and plant derived products such as enhanced extract, fraction as well as pure compounds.
15.	Quality Phytochemicals	2000	Song Gao	New Jersey, USA	http://www.qualityphytochemicalsllc.com	Purified Phyto-chemicals or natural compounds
16.	Selleck Chemicals	2009	Dean Henry	Texas, United States	http://www.selleckchem.com	Purified natural products for research
17.	Sequoia Sciences	1999	Gary Eldridge & Mark O'Neil-Johnson	Seattle, Washington, USA	https://sequoiasciences.com sequoia@sequoiasciences.com	Drug-like compounds isolated from plants and other sources that aid in developing new drug targets for various diseases.
18.	Specs	1987		Kluyverweg, Netherlands	https://www.specs.net/	Natural products and byproducts from plants, fungi, bacteria, and marine organisms.
19.	Target Molecule Corp.				https://www.targetmol.com/	Purified natural compounds like Alkaloids, Flavonoids, Phenols etc, obtained from plant, animal and microbes.
20.	TimTec	1995	Dr. Murat Niyazymbetov	Newark, Delaware, USA	www.timtec.net	Compounds obtained from plants, bacteria, fungus, and animal sources.

Abbreviations: NCI, national cancer institute; NIH, national institute of health

institutes to produce a huge collection of natural extract which was originally designed to assist the discovery of anti-cancer drugs [10]. Today, although it is non-essential to combine several organisms to acquire new bioactive molecules, likelihoods of accomplishment are meticulously allied with the choice of unambiguous assortments and the extent of unique samples. This can be achieved by maximizing the variety of natural products in bioactive compound libraries organized from microbes, plants, marine organisms and additional materials, provided the biodiversity, state, region or taxonomic group is well characterized.

Assembly of natural extracts with biomedical applications could be restricted to a specified group or *genera* or particularly designed bioactive compounds; for instance, allowing the study of a sequence with the help of given scaffold types [11]. In some laboratories, assessments of a few hundred natural biomedical compounds in different sets are common in exhaustive studies, also these small collections are easy to manage [12]. Nevertheless, to enhance the likelihoods of obtaining novel natural bioactive compounds from composite mixtures, the speedy initial step such as high-performance liquid chromatography (HPLC) or solid phase extraction is required for the enriching of natural raw materials [13]. Such operations can perk up further bio-chemical screening procedures. Additionally, these compounds can be identified faster with high throughput screenings, using emerging state-of-the-art mass spectrometry and omics technologies [14].

3. Bioactive compounds for cancer and Omics approaches

Biological research has revolutionized with the commencement of omics era, that enables the expansion of high-end technology for the attainment and analysis of the large available datasets [15]. This all-inclusive approach is poised to determine and comprehend novel patterns and possesses a wide range of applications that are important in the arena of biotechnology.

Over past decade, expanding public and private inquisitiveness and investments in cancer research have augmented the prospects to generate evidence, and assemble large quantities of data to comprehend cell death processes under biological conditions [16]. Multi-omics approach for cancer treatment integrates multiple omics methods (such as genomics, transcriptomics, proteomics and metabolomics) for the production and analysis of huge chemotherapeutic natural product data [17]. Out of many, the utmost promising breakthrough of omics biotechnology is the extensive identification of plant-based bioactive compounds.

For the past several years, biologists have been using genomics in almost all aspects of life, and the number of genomic resources in phytochemicals has now become progressively more significant [18]. One of the foremost restrictions on the usage of these resources for the development of cancer related techniques is the scarcity of complete online databases where complete information related to plant genome is available [19]. Comparative genomics of different groups of plants can help in identifying the genetic basis of desirable traits; in addition, the genetic composition accountable for the specified phenotype will facilitate the practice of genetic engineering in improving the cancer therapeutic sector [20].

Numerous transcriptomes have been vigilantly interpreted and considered, providing an exceptional stance into the miscellaneous transcription-related processes operating in plants [21]. Taking into consideration the great therapeutic significance of phytochemicals, the transcriptomic scheme has been applied in order to disclose the genetic pathways liable for pathogenesis and provide viable therapeutic targets in different type of cancers [21]. Both transcriptomics and genomics alone are insufficient to comprehend the multifaceted biology of phytochemicals and ought to be harmonized through proteomic and metabolomics approaches, *inter alia*.

Proteomics explains the processes involved in biological mechanism and their functions and also provides information of protein including the pre- and post-transcriptional modifications, as well as protein-

protein interactions [22]. Plant metabolites also have unique therapeutic properties and stages of their biosynthesis dependent on genetics and environmental changes [23]. A full-scale investigation of plant metabolites is described as metabolomics [24]. Almost all plants with medicinal values yield a scarce variety of secondary active metabolites that are different from compounds documented in other plants, as they have distinct metabolic patterns that are highly interconnected with the distinctive features of their landscapes, which contain an example of continuous variation in physical parameters such as light, pressure, nutrients etc. [25]. The desired plant metabolites having great interest in cancer drug discovery are acknowledged and investigated in a targeted way, and usually, the quantity of metabolites isolated is miniscule. For high throughput metabolite profiling, ancillary techniques such as mass spectrometry (MS) and nuclear magnetic resonance (NMR) are essential [26].

In recent years, omics approaches have been endowed with new opportunities to recognize and illustrate high-value bioactive compounds from plant origin that are effective against OMCs [27]. However, exploring the discovery of therapeutic phytochemicals, involves the use of poorly annotated data bases, inadequate reference datasets, and a dearth of committed software. Regardless of declining expenses on omics approaches, the datasets accessible are still insufficient for relative tools to work proficiently and additional investments into the gathering of data from plant species is crucial. Therefore, integrative multi-omics approaches will eventually accelerate the discovery of reliable cancer drugs in various plant species [28].

The bioactive elements entrenched in plants have been the precursors of modern research into natural product research and are a major source of drug carrier molecules [29]. Synergistic integration of bioinformatics and omics data (transcriptomics, physiogenomics, proteomics and metabolomics) could be utilized to explore and evaluate natural compound's mechanisms, in a more precise way. Emerging omics technologies allow the multiplexing and parallel processing of proteomics and metabolic data [30]. In addition, an evidence-based integration of modern biomedical tools and traditional medicine practices can remarkably transform new drug development strategies, leading to precision and personalized medicine for the treatment of several type of cancers including OMCs. Bioinformatics provides information on diseases, toxicity issues, treatment factors, etc. [31].

4. Biomedical applications of bioactive compounds

It is well-known that in primeval times, people used nature to accomplish their elementary requirements including usage of natural products as medicines for various diseases. Indeed, an archeological study has provided information that about 50,000 years ago, Neanderthals were acquainted with therapeutic values of medicinal plants and used them for remedial purposes [32]. Apart from ancient uses, the most primitive written chronicles of medicinal plants have been found in Mesopotamia, 2600 BCE, unfolding the practice of using some gymnosperms like cedar besides medicinal oils intended for treating certain conditions like cough, inflammation and fever [33]. Interestingly, the people of this region as well other parts of the world still practice using these plant parts and their ingredients as medicines [34]. Nowadays, approximately 70–95% of the residents in developing countries of Asia and Africa uses traditional medicine for the treatment of various ailments [35].

Phytochemicals and plant-derived products are hopeful options meant for improving clinical effectiveness of cancer patients and reducing adverse reactions [36]. Most of these phytochemicals are natural active chemicals carrying high anticancer properties [37]. The development of high-quality anticancer drugs with enviable effects commences with testing of natural extracts to detect antitumor activity trailed by isolation of active phytochemicals following *in vitro* and *in vivo* results as shown in Fig. 2 [38]. Various bioactive compounds extracted from different regions of plants have been identified for the

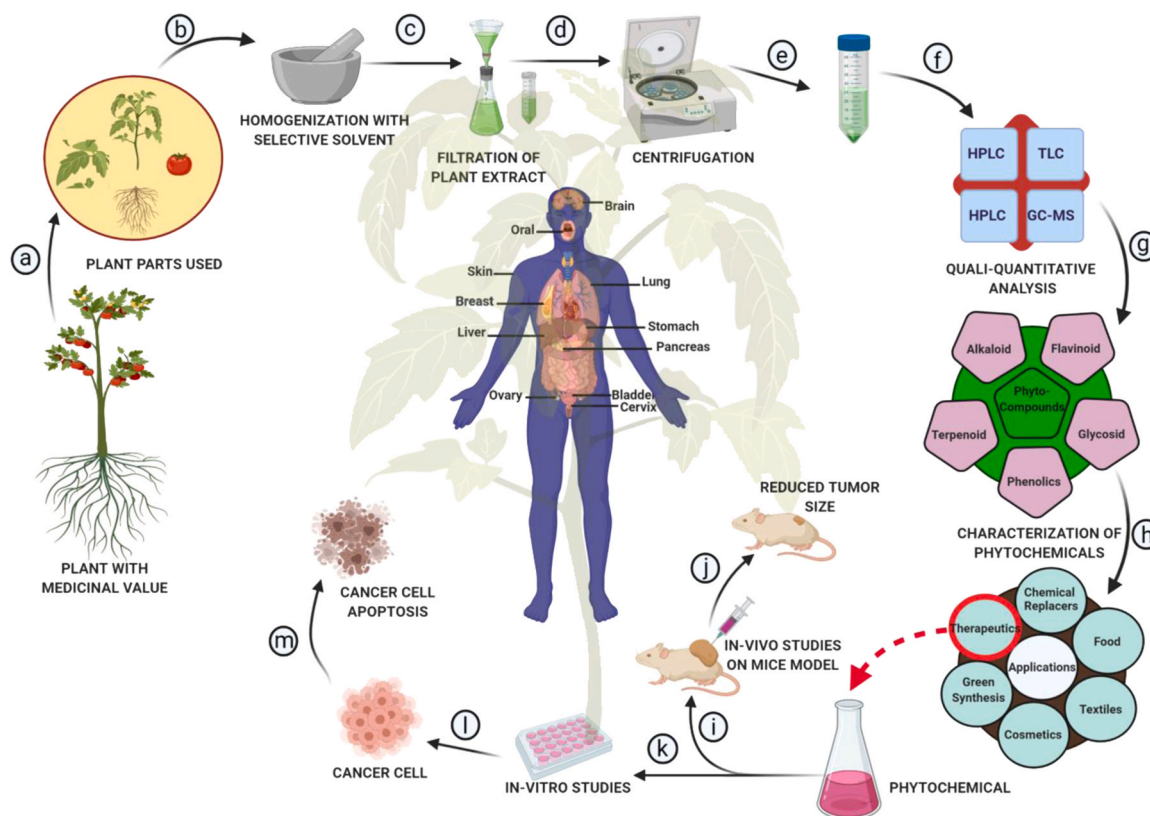


Fig. 2. Extraction of bioactive phyto-chemicals and their applications. Abbreviations: HPLC, high-performance liquid chromatography; TLC, thin layer chromatography; GC-MS, gas chromatography mass spectrometry. Explanatory notes: a. Selection of plant with medicinal properties and isolation of plant parts to be used for extraction of bioactive compound b. Homogenization of plant part is carried out by using appropriate solvents. c-d. Homogenized extracts are subjected to filtration and centrifugation to obtain pure plant extracts e-f. Qualitative cum quantitative analysis of the Phyto-chemicals through various techniques such as HPLC, TLC, GC-MS etc. g. Characterization of obtained Phyto-chemicals into different groups h. Applications of Phyto-chemicals i-m. In-vivo (on xenografts) and in-vitro studies to check therapeutic potential of bioactive compounds obtained from plants (Created with www.BioRender.com).

treatment of human pathologies (Table 2). As depicted by many studies inhibition of COX-2 by Apigenin results in anti-inflammatory activity [39]. Also, use of Berberin hinders mitochondrial functions of target cells and can be used as a therapeutic prospective against numerous disorders [40]. Cryptolepine extracted from *Cryptolepis sanguinolenta* inhibits DNA synthesis showing cyto-toxic activity against malarial parasite [41]. Curcumin and Epigallocatechin-3- gallate isolated from different source plants are well known for their anti-cancer properties [42,43]. Gossypol an extract isolated from *Gossypium* acts as an ant fertility agent by thwarting enzymes that are involved in energy providing metabolic reactions in spermatocytes [44]. Antiproliferative, antioxidant and cytotoxic activity of Linalool, Lycopene and Piperine include inhibition of dehydrogenase enzyme, quenching singlet oxygen and inhibition of the activity of EGFR tyrosine kinase enzyme. respectively [45,46]. Quercetin works as an anti cancer agent by block the PI3K / AKT pathway leading to a reduction in the expression of anti-apoptotic protein, thus acting as an anti cancer agent [47]. Apart from these, bioactive compounds such as Resveratrol, Sesquiterpene Coumarins, Sesquiterpenoids, Ursolic acid, Withaferin A, Apomorphine etc., have been validated for their efficacy in various diseases including cancer [53]. Arteether is used as an antimalarial agent as it inhibit the nutrient flow to erythrocytic stage of *Plasmodium falciparum* by modifying properties of membranes [54]. Galantamine, an alkaloid extracted from *Galanthus woronowii* is exploited for the treatment of Alzheimer as it escalates acetylcholine neurotransmission [55]. *Callistemon citrinus* extracts causes obstruction in the catabolic pathway of tyrosine that leads to a decline in the accumulation of venomous metabolites in Hepatorenal tyrosinemia [56]. It has been observed that Paclitaxel mediates anti cancer activity through endorsing arrest of mitosis and cell

death [57]. Leaf and root extracts of *Atropa belladonna* have been found effective against asthma and chronic obstructive pulmonary disease by acting on specific muscarinic receptors located in the respiratory tract [58].

Many of these plants are endemic to the African and Asia continent and their use to ameliorate the increasing burdens of OMC cancers. In these regions that constitute a high percentage of the global low- and middle-income countries, should be explored.

5. Bioactive plant compounds for the management of OMC cancers

Cancer is major causes of death across the globe and owing its ubiquity the recognition of new anti-cancer remedies is imperative. The previous century has made prodigious strides in plant research and phytochemicals field, through admittance of numerous anti-cancer natural compounds already included in treatment practices [59]. A list of commercially accessible plant derived bioactive compounds for treating numerous cancers has been provided in Table 3, along with details like their source plant, plant family, plant part used for extraction of compound, type of studies (in-vitro/in-vivo) performed so far.

Several thriving anti-cancer drugs used in the therapeutics, revealed to be very efficient, are invented as natural products like microbes, plants and marine organisms [60]. Current developments in proteomics along with metabolomics have proven imperative for identification of novel therapeutic targets that proffer innovative therapeutic ideas. Most of the molecules with medicinal properties which were identified in the late 90 s and early 2000s, are natural products or their derivatives, utterly substituting antitumor agents [61]. In this perception, natural

Table 2
Biomedical Applications of Plant Derived Bioactive Compounds.

	Bioactive compound/ Natural product	Category/ Chemical nature	Origin/source plant	Plant family	Plant part used	Mechanism of action	Biomedical application	References
1.	Apigenin	Flavonoid	<i>Salvia officinalis</i>	Lamiaceae	Whole plant	Inhibition of COX-2 results in anti-inflammatory activity.	Antioxidant, anti-inflammatory, anti-mutagenic, antibacterial and antiviral properties	[39]
2.	Berberin	Alkaloid	<i>Coptis chinensis</i>	Ranunculaceae	Roots	Inhibit the function of mitochondria, prompt glycolysis and commencement of AMPK pathway.	Therapeutic prospective against numerous disorders (Cardiac disorders, Neurological disorders and Diabetes) Antibacterial, Anti-inflammatory, Cytotoxic activity	[40]
3.	Casticin	Flavonoids	<i>Artemisia annua</i>	Asteraceae	Fruits	Expression of Bax protein is up regulated and Bcl-2 protein expression is down-regulated that results in arrest of the cell cycle at G2/M phase and induce apoptosis.	Anti-proliferation, Inhibit self-renewal of hepatic stem cells, Anti-inflammatory	[76]
4.	Cryptolepine	Alkaloids	<i>Cryptolepis sanguinolenta</i>	Apocynaceae	Roots	Cyto-toxic activity of drug is as a result of inhibition of DNA synthesis.	Cyto-toxic activity, Antimalarial, Antiplasmodial activity	[41]
5.	Curcumin	Polyphenols	<i>Curcuma longa</i>	Zingiberaceae	Rhizome	Initiation of apoptosis Obstruction of nuclear factor-kappa B (NF-κB) instigation Downregulation of numerous pro-inflammatory cytokines (IL-8, IL-6, and TNFα)	Anti-cancer Anti-inflammation Anti-angiogenic Antioxidant, and anti-mutagenic effects	[42]
6.	Epigallocatechin-3-gallate (EGCG)	Polyphenols	<i>Camellia sinensis</i>	Theaceae	Dried leaves	Antioxidan Drug resistance reversal	Antioxidative activity Anti-cancer DNA-protective Neuroprotective effect Anti-HIV	[43]
7.	Gingerol	Guaiacols	<i>Zingiber officinale</i>	Zingiberaceae	Rhizome	Inhibition of NF-κB signaling pathway	Antioxidant and anti-inflammatory properties	[77]
8.	Gossypol	Polyphenolic Aldehyde	<i>Gossypium</i>	Malvaceae	Whole plant	It prohibits production and motility of male gametes It acts as an antifertility agent by thwarting enzymes that are involved in energy providing metabolic reactions (in sperm producing cells and spermatozoa)	Antifertility, antiviral, anticancer, antioxidant, antitrypanosomal, antimicrobial, and antimalarial activities	[44]
9.	Linalool	Acyclic Monoterpenoid	<i>Hyptis crenata</i>	Lamiaceae	Leaves	Inhibit the activity of the respiratory chain dehydrogenase enzyme.	Antiproliferative antioxidant and cytotoxic and effects, antibacterial and insecticidal activity, ulcer healing properties.	[45]
10.	Lycopene	Carotenoid	<i>Solanum lycopersicum</i>	Solanaceae	Fruit	Lycopene is a powerful quencher of singlet oxygen which is an active form of oxygen, signifying its role as a more effective antioxidant.	Antioxidant activity	[46]
11.	Piperine	Alkaloid	<i>Piper nigrum</i>	Piperaceae	Fruit	Inhibit the activity of EGFR tyrosine kinase enzyme.	Reduction of insulin-resistance, anti-inflammatory effects	[46]
12.	Platycodon saponin	Amphipathic Glycosides	<i>Platycodon grandiflorum</i>	Campanulaceae	Roots	Functional inhibition of NF-κB and PI3K / AKT and MAPK signaling pathways.	Anti-cancer cytotoxic cells, Antiviral activity neuroprotective activity, Cholesterol lowering properties	[78]
13.	Quercetin	Polyphenolic Flavonoid	<i>Capparis spinosa</i>	Capparaceae	Leaves	Blockage of the PI3K / AKT pathway leading to a reduction in the expression of anti-apoptotic protein Bcl-w.	Antioxidant and hepatoprotective effects, Anti-cancer	[47]
14.	Resveratrol	Polyphenols	<i>Vitis vinifera Linnaeus</i> (grapes)	Vitaceae	Fruit	cell survival signals inhibitor, p53 activation Apoptosis Inhibitor	Anti-cancer Anti-oxidant Anti-inflammation Cardioprotection	[48]
15.	Sesquiterpene Coumarins	Terpene	<i>Mikania spp</i>	Asteraceae	Leaves and aerial parts	Restrain the exflagellation of Plasmodial male microgamete by causing DNA damage in cells undergoing replication.	Cytotoxic activity Antibacterial activity anti-inflammatory and antitumor activity.	[49]

(continued on next page)

Table 2 (continued)

	Bioactive compound/ Natural product	Category/ Chemical nature	Origin/source plant	Plant family	Plant part used	Mechanism of action	Biomedical application	References
16.	Sesquiterpenoids	Terpene	<i>Inula linearifolia</i>	Asteraceae	Aerial parts	Act in response to the active groups accessible in proteins and enzymes, particularly the thiol group.	Induction of apoptosis, Cytotoxic activity; Inhibition of breast cancer cell growth	[50]
17.	Ursolic acid	Triterpenoids	<i>Ocimum basilicum</i>	Lamiaceae	Whole plant	significant action in glucose homeostasis, reduces diabetic blood glucose levels, Improves the glucose and insulin tolerance, surges insulin synthesis and release from pancreas and prevents protein glycation	Antiviral activity Antiproliferative activity	[51]
18.	Withaferin A	Withanolide	<i>Withania sonnifera</i>	Solanaceae	Leaves	Drug interacts with C-terminus of Hsp90, thus provoking degradation of heat shock proteins.	Antioxidant activity, Anti-proliferative	[52]
19.	Apomorphine	Dopamine receptor agonist	<i>Papaver sonniferum</i>	Papaveraceae	Unripe capsules	Drug acts by enhancing motor function and vasodilation by exciting dopamine receptors (D2) in hypothalamus, pituitary gland, and blood vessels.	Parkinson's disease	[53]
20.	Arteether	Sesquiterpene trioxane lactone	<i>Artemisia annua</i>	Asteraceae	Leaves	Inhibit the nutrient flow to erythrocytic stage of parasite (<i>P. falciparum</i>) by modifying properties of membranes.	Antimalarial	[54]
21.	Galantamine	Amaryllidaceae alkaloid	<i>Galanthus woronowii</i>	Amaryllidaceae	Bulb	Esclates acetylcholine neurotransmission	Alzheimer	[55]
22.	Nitisinone	Mesotriene	<i>Callistemon citrinus</i>	Myrtaceae	Roots	It causes obstruction in the catabolic pathway of tyrosine that leads to a decline in the accumulation of venomous metabolites in HT-1.	Hepatorenal tyrosinemia Anti-cancer activity	[56]
23.	Paclitaxel	Taxane diterpene	<i>Taxus brevifolia Nutt.</i>	Taxaceae	Bark	Alleviates microtubules and diminish their dynamicity. Endorse arrest of mitosis and cell death.	Anti-cancer activity	[57]
24.	Tiotropium	Muscarinic receptor antagonist	<i>Atropa belladonna</i>	Solanaceae	leaves and roots	Acts on specific muscarinic receptors located in the respiratory tract to cause relaxation of smooth muscles and dilation of bronchioles.	Asthma and COPD (<i>Chronic obstructive pulmonary disease</i>)	[58]

Abbreviations: COX-2, cyclooxygenase-2; COPD, chronic obstructive pulmonary disease; HT-1, Hepatorenal tyrosinemia-1; dopamine receptors (D2); Hsp90, heat shock protein 90; PI3K, Phosphoinositide 3-kinase; PKB, protein kinase B; MAPK, mitogen-activated protein kinase; NF- κ B, Nuclear factor kappa B; IL-8, Interleukin 8; IL-6, Interleukin 6; TNF α , tumor necrosis factor-alpha; AMPK, AMP-activated protein kinase; EGFR, epidermal growth factor receptor.

compounds have been known as the widely abundant and efficient source of new anti-cancer drugs [62].

Various scientific evidences signify the importance of a number of phyto-chemicals in cancer therapeutics [63]. Phyto-chemicals which have been established carrying anti-cancer activity through have a number of consistent mechanisms for slowing down the cancer progression by declining survival and proliferation of malignant cells, removing free radicals, and reducing the angiogenesis of tumors [64]. They work with a wide range of complex mechanisms of action in many signaling pathways including membrane receptors, oncoproteins or tumour-suppressor proteins, transcription factors, microRNAs and caspases [65]. A few therapeutic bioactive compounds have been identified (Table 4) that has shown therapeutic potential in the management of OMCs. For instance Epicatechingallate derived from leaves of *Camelia sinensis*, Gingerol isolated from *Zingiber officinale* roots and Benzylisothiocyanate extracted from seeds of *Moringa oleifera* has been tested for their efficacy against oral cancer [66–68].

Epicatechingallate mediates anti cancerous activities such as inhibition of migration, invasion, angiogenesis, and promotion of apoptosis through modulating the production of reactive oxygen species (ROS), inhibiting the pathway associated with nuclear factor- κ B signaling, promoting the modifications at epigenetic level through regulating

acetylation of histones and inhibiting DNA methyltransferase activity [69]. Gingerol mediates anti cancerous activity in oral cancer by inducing apoptosis and arresting cell cycle [70]. Oral cancer when treated with Benzylisothiocyanate there is a rapid production in reactive oxygen species that promote DNA damage, which along with redox stress activates p21 and p53, resulting in cell cycle arrest [71]. Resveratrol, solasonine, curcumin and several other bioactive compounds are found to be effective against skin cancer. There is a paucity of the use of plant bioactive compound and libraries in the management of OMCs. Hence, there is a pressing need to employ both in-silico and in-vitro approaches to develop compound libraries that would be amenable for the identification of useful therapeutic targets for OMC cancers in the era of precision medicine.

6. Conclusion and future perspectives

Cancer, a foremost public health issue, has a profound influence across the globe and influencing both developed as well developing countries [72]. Given the severity of cancer, its handling and treatment has been an enduring struggle with insignificant accomplishment. Presently available cancer treatment includes surgical removal following radiation treatment for a cancer mass, which is often followed

Table 3
List of commercially accessible plant derived compounds/drugs for treating numerous cancers.

Cancer type	Plant Family	Plant	Plant Part Used	Phyto Compound	Type of studies	References	
1. Bladder Cancer	Theaceae	<i>Camellia sinensis</i>	Leaves	Epicatechingallate	Both in vitro and in vivo	[79]	
	Zingiberaceae	<i>Zingiber officinale</i>	Roots	Gingerol	Both in vitro and in vivo	[80]	
2. Blood Cancer	Plumbaginaceae	<i>Plumbago zeylanica</i>	Roots	Plumbagin	In vitro	[81]	
	Ranunculaceae	<i>Clematis manshrica</i>	Flower and Leaves	Benzoquinone	In vivo	[82]	
	Plumbaginaceae	<i>Plumbago zeylanica</i>	Roots	Plumbagin	In vitro	[83]	
	Theaceae	<i>Camellia sinensis</i>	Leaves	Epigallocatechin gallate	In vivo	[84]	
3. Brain Cancer	Zingiberaceae	<i>Curcuma longa</i>	Rhizome	curcumin	In vitro	[85]	
	Vitaceae	<i>Vitis vinifera</i>	Leaves	Resveratrol	Both in vitro and in vivo	[86]	
4. Breast Cancer	Theaceae	<i>Camellia sinensis</i>	Leaves	Epigallocatechin gallate		[87]	
	Cannabinaceae	<i>Cannabis sativa</i>	Leaves	Cannabinoid	Both in vitro and in vivo	[88]	
	Rhamnaceae	<i>Ziziphus spina-christi</i>	Flowers and Leaves	Doxorubicin	In vivo	[89]	
	Apiaceae	<i>Centella asiatica</i>	Leaves	Asiatic acid	In vitro	[90]	
	Theaceae	<i>Camelia sinesis</i>	Leaves	Epicatechingallate, picatechin, epigallocatechin	Both in vitro and in vivo	[91]	
	Araliaceae	<i>Panax ginseng</i>	Leaves	Panaxadiol	Both in vitro and in vivo	[92]	
	Zingiberaceae	<i>Zingiber officinale</i>	Roots	Gingerol	In vitro	[93]	
	Zygophyllaceae	<i>Peganum harmala</i>	Roots	Harmine	In vitro	[94]	
	Apiaceae	<i>Centella asiatica</i>	Whole plant	Tamoxifen	Both in vitro and in vivo	[95]	
	Rhamnaceae	<i>Ziziphus jujuba</i>	Fruits, Seeds and Leaves	Linoleic acid	In vivo	[96]	
	Lamiaceae	<i>Ocimum sanctum</i>	Leaves	Eugenol	Both in vitro and in vivo	[97]	
	Cucurbitaceae	<i>Momordica charantia</i>	Leaves and Roots	Charantin	In vitro	[98]	
	5. Cervical Cancer	Amaryllidaceae	<i>Allium sativum</i>	Buds and Leaves	Allicin	In vivo	[99]
		Rhamnaceae	<i>Ziziphus mauritiana</i>	Leaves, Bark and Fruits	Methyl stearate	In vitro	[100]
Crassulaceae		<i>Bryophyllum pinnatum</i>	Leaves	Bryophyllin	In vitro	[101]	
Solanaceae		<i>Withania somnifera</i>	Roots, Stem and Leaves	5-Fluorouracil	In vitro	[102]	
Zingiberaceae		<i>Zingiber officinale</i>	Roots	Gingerol	In vitro as well as In vivo	[103]	
6. Colon Cancer	Araliaceae	<i>Panax ginseng</i>	Leaves	Panaxadiol	In vitro	[104]	
	Ginkgoaceae	<i>Ginkgo biloba</i>	Leaves	Bilobalide	In vitro	[105]	
	Vitaceae	<i>Vitis vinifera</i>	Seeds extract and Fruits	Procyanidins	In vivo	[106]	
	Zingiberaceae	<i>Curcuma longa</i>	Rhizomes	Curcumin, ascorbic acid	Both in vitro and in vivo	[107]	
	Moringaceae	<i>Moringa oleifera</i>	Seeds	benzylisothiocyanate, 4-benzylisothiocyanate	In vitro	[108]	
	Passifloraceae	<i>Passiflora caerulea</i>	Flowers	Chrysin	In vitro	[109]	
	Solanaceae	<i>Capsicum annum</i>	Pepper	Luteolin	In vitro	[110]	
	Dioscoreales	<i>Dioscorea colletti</i>	Rhizomes	Dioscin	In vitro	[111]	
	Schizophyllaceae	<i>Schizophyllum commune</i>	Fruiting bodies	Quecertain	Both in vitro and in vivo	[112]	
	8. Head and Neck Cancer	Betulaceae	<i>Betula Sp.</i>	Leaves	Betulinic acid	In vitro	[113]
Zingiberaceae		<i>Curcuma longa</i>	Rhizomes	Curcumin	Both in vitro and in vivo	[114]	
9. Leukemia	Rhamnaceae	<i>Ziziphus spina-christi</i>	Flowers and Leaves	Doxorubicin	Both in vitro and in vivo	[115]	
	Asteraceae	<i>Xanthium strumarium</i>	Fruits	Xanthatin	In vitro	[116]	
10. Liver Cancer	Zingiberaceae	<i>Curcuma longa</i>	Rhizomes	Curcumin, ascorbic acid	In vitro	[117]	
	Polygonaceae	<i>Fagopyrum sculentum</i>	Seeds	Buckwheat inhibitor-1protein	In vitro	[118]	
	Convolvulaceae	<i>Ipomoea batata</i>	Roots	Trypsin inhibitor protein	In vitro	[119]	
	Asteraceae	<i>Xanthium strumarium</i>	Fruit s	Promyelocytic Xanthatin	In vitro	[120]	
	Dioscoreales	<i>Dioscorea colletti</i>	Rhizomes	Dioscin	In vitro	[121]	
	Malvaceae	<i>Hibiscus mutabilis</i>	Pepper	Lectin	In vitro	[122]	
10. Liver Cancer	Polygonaceae	<i>Polygonum cuspidatum</i>	Whole plant	Resveratrol	In vitro	[123]	
	Lamiaceae	<i>Ocimum sanctum</i>	Leaves	Eugenol, orientin, vicenin	Both in vitro and in vivo	[124]	
	Solanaceae	<i>Solanum nigrum</i>	Leaves	Solasonine	In vitro	[125]	
	Asclepiadaceae	<i>Asclepias curassavica</i>	Aerial parts	Cardenolides	In vitro	[126]	

(continued on next page)

Table 3 (continued)

Cancer type	Plant Family	Plant	Plant Part Used	Phyto Compound	Type of studies	References
	Nelumbonaceae	<i>Nelumbo nucifera</i>	Embryos	Neferine	In vitro	[127]
	Theaceae	<i>Camelia sinensis</i>	Leaves	Epicatechingallate	In vitro	[128]
	Moringaceae	<i>Moringa oleifera</i>	Seeds	Benzylisothiocyanate	In vitro	[129]
	Rhamnaceae	<i>Ziziphus spina-christi</i>	Flowers and Leaves	Doxorubicin	In vivo	[130]
	Iridaceae	<i>Saffron crocus</i>	Dry stigmas	Saffron	Both in vitro and in vivo	[131]
	Solanaceae	<i>Solanum nigrum</i>	Leaves	Solasonine	In vitro	[132]
	Theaceae	<i>Camellia sinensis</i>	Leaves	Theabrownin	In vivo	[133]
	Liliaceae	<i>Crocus sativus</i>	Dry stigmas	Crocin	In vivo	[134]
	Podophyllaceae	<i>Podophyllum peltatum</i>	Leaves	Podophyllotoxin	In vitro	[135]
11. Lymphoma	Rhamnaceae	<i>Ziziphus rugosa</i>	Pericarp and Seeds	Betulinic acid	In vivo	[136]
12. Nasopharyngeal Carcinoma	Fabaceae	<i>Phaseolus vulgaris</i>	Seeds	Lectin	In vitro	[137]
	Zingiberaceae	<i>Curcuma longa</i>	Rhizomes	Curcumin	In vitro	[138]
13. Ovarian Cancer	Annonaceae	<i>Annona squamosa</i>	Seeds	Bullatacin	In vitro	[139]
	Zingiberaceae	<i>Zingiber officinale</i>	Rhizomes	6-Shogaol	In vitro	[140]
	Amaryllidaceae	<i>Allium sativum</i>	Buds and Leaves	Allicin	In vitro	[141]
14. Pancreatic Cancer	Amaryllidaceae	<i>Allium sativum</i>	Buds and Leaves	Allicin	In vitro	[142]
	Zingiberaceae	<i>Zingiber officinale</i>	Roots	Gingerol	Both in vitro and in vivo	[143]
	Theaceae	<i>Camelia sinensis</i>	Leaves	Epicatechingallate	Both in vitro and in vivo	[144]
15. Prostate Cancer	Cannabinaceae	<i>Cannabis sativa</i>	Leaves	Cannabinoid	Both in vitro and in vivo	[145]
	Theaceae	<i>Camellia sinensis</i>	Leaves	Epigallocatechin gallate	In vivo	[146]
	Solanaceae	<i>Solanum lycopersicum</i>	Fruits	Lycopene	In vivo	[147]
	Fabaceae	<i>Cicer arietinum</i>	Seeds	Bowman-Birk-type protease	In vitro	[148]
	Polygonaceae	<i>Polygonum cuspidatum</i>	Whole plant	Resveratrol	In vivo	[149]
16. Stomach Cancer	Asphodelaceae	<i>Aloe barbadensis</i>	Leaves	Emodin	In vivo	[150]

Table 4

List of commercially available plant derived compounds/drugs for treating oral mucocutaneous cancers (OMCs) and Skin Cancers.

Cancer type	Plant Family	Plant	Plant Part Used	Phyto Compound	Type of studies	References
1. Oral Cancer	Theaceae	<i>Camelia sinensis</i>	Leaves	Epicatechingallate, picatechin, epigallocatechin	Both in vitro and in vivo	[66]
	Zingiberaceae	<i>Zingiber officinale</i>	Roots	Gingerol	In vitro	[67]
	Moringaceae	<i>Moringa oleifera</i>	Seeds	Benzylisothiocyanate	In vitro	[68]
2. Skin Cancer	Polygonaceae	<i>Polygonum cuspidatum</i>	Whole plant	Resveratrol	In vitro	[151]
	Apocynaceae	<i>Carissa spinarum</i>	Fruits	Alkaloids, saponins, tannins, flavonoids	In vitro	[152]
	Solanaceae	<i>Solanum nigrum</i>	Leaves	Solasonine	In vitro	[153]
	Plumbaginaceae	<i>Plumbago zeylanica</i>	Roots	Plumbagin	In vitro	[154]
	Zingiberaceae	<i>Curcuma longa</i>	Rhizomes	Curcumin	In vitro	[155]

by treatment with the use of preservative chemicals [73]. Existing chemotherapeutic treatments include DNA-interactive compounds like Cisplatin, antimetabolites (e.g. Methotrexate), hormones, anti-tubulins including taxanes, in addition to cell identification agents [74]. The main disadvantage of chemotherapy is the cancer relapse, drug resistance, along with lethal effects on unintended tissues obstructing the usage of anti-cancer drugs, consequently affecting the patient's survival and life quality [75]. To overcome current medical problems, we must look for novel and effective anticancer agents carrying improved efficacy and minimum side effects. We have discussed in this review the potential for the use of bioactive plant compounds as potential complementary approach to the management of OMCs which have high burdens in India and Africa. We have also highlighted the benefit of the use of high throughput omics technique to develop medicinal and anti-OMC cancerous compound libraries from widely endemic plants in these regions. Furthermore, a catalog of bioactive compounds libraries used for various cancers, including OMCs has been provided. It is hoped that bioactive compounds would be further explored for the management of oral and maxillofacial diseases, particularly, OMCs.

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Henry A. Adeola: Data curation, Writing - original draft preparation; **Afsareen Bano:** Data curation, Writing - original draft preparation; **Ravina Vats:** Writing - original draft preparation; **Amit Vashishtha:** Writing - review & editing; **Deepika Verma:** Writing - review & editing; **Deepak Kaushik:** Figure creation and Editing; **Vineet Mittal:** Figure creation and Editing; **Md. Habibur Rahman:** Writing - review & editing; **Agnieszka Najda:** Writing - review & editing; **Ghadeer M. Albadrani:** Writing - review & editing; **Amany A. Sayed:** Writing - review & editing; **Sameh M. Farouk:** Writing - review & editing; **Emad H. M. Hassanein:** Writing - review & editing, Validation; **Muhamad Furqan Akhtar:** Writing - review & editing, Validation; **Ammara Saleem:** Writing - review & editing; **Mohamed M. Abdel-Daim:** Conceptualization, Supervision; **Rashmi Bhardwaj:** Conceptualization,

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Conflict of interest statement

The authors declare that they have no competing interests.

Data Availability

Data sharing is not applicable to this article as no datasets were generated or analysed during the current study.

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References

- [1] N. Van Hilten, F. Chevillard, P. Kolb, Virtual compound libraries in computer-assisted drug discovery, *J. Chem. Inf. Model.* 59 (2019) 644–651, <https://doi.org/10.1021/acs.jcim.8b00737>.
- [2] M. Stevens, S. Abdeen, N. Salim, A.M. Ray, A. Washburn, S. Chitre, J. Sivinski, Y. Park, Q.Q. Hoang, E. Chapman, S.M. Johnson, HSP60/10 chaperonin systems are inhibited by a variety of approved drugs, natural products, and known bioactive molecules, *Bioorg. Med. Chem. Lett.* 29 (2019) 1106–1112, <https://doi.org/10.1016/j.bmcl.2019.02.028>.
- [3] A. de la Vega de Leon, E. Lounkine, M. Vogt, J. Bajorath, Design of diverse and focused compound libraries. Tutorials in Chemoinformatics, John Wiley & Sons, Ltd, Chichester, UK, 2017, pp. 83–101, <https://doi.org/10.1002/9781119161110.ch5>.
- [4] E.K. Davison, M.A. Brimble, Natural product derived privileged scaffolds in drug discovery, *Curr. Opin. Chem. Biol.* 52 (2019) 1–8, <https://doi.org/10.1016/j.cbpa.2018.12.007>.
- [5] R. Friedman Ohana, S. Levin, M.G. Wood, K. Zimmerman, M.L. Dart, M. K. Schwinn, T.A. Kirkland, R. Hurst, H.T. Uyeda, L.P. Encell, K.V. Wood, Improved deconvolution of protein targets for bioactive compounds using a palladium cleavable chloroalkane capture tag, *ACS Chem. Biol.* 11 (2016) 2608–2617, <https://doi.org/10.1021/acscchembio.6b00408>.
- [6] A. Quintavalla, Spirolactones: recent advances in natural products, bioactive compounds and synthetic strategies, *Curr. Med. Chem.* 25 (2018) 917–962, <https://doi.org/10.2174/0929867324666171106162259>.
- [7] M. Koromina, M.T. Pandi, G.P. Patrinos, Rethinking drug repositioning and development with artificial intelligence, machine learning, and omics, *OMICS A J. Integr. Biol.* 23 (2019) 539–548, <https://doi.org/10.1089/omi.2019.0151>.
- [8] P.E. Saw, S. Lee, S. Jon, Naturally occurring bioactive compound-derived nanoparticles for biomedical applications, *Adv. Ther.* 2 (2019), 1800146, <https://doi.org/10.1002/adtp.201800146>.
- [9] J.L. Wolfender, M. Litaudon, D. Touboul, E.F. Queiroz, Innovative omics-based approaches for prioritisation and targeted isolation of natural products—new strategies for drug discovery, *Nat. Prod. Rep.* 36 (2019) 855–868, <https://doi.org/10.1039/c9np00004f>.
- [10] B.A.P. Wilson, C.C. Thornburg, C.J. Henrich, T. Grkovic, B.R. O'Keefe, Creating and screening natural product libraries, *Nat. Prod. Rep.* 37 (2020) 893–918, <https://doi.org/10.1039/c9np00068b>.
- [11] A.S.A. El-Sayed, M.T. El Sayed, A. Rady, N. Zein, G. Enan, A. Shindia, S. El-Hefnawy, M. Sitohy, B. Sitohy, Exploiting the biosynthetic potency of taxol from fungal endophytes of conifers plants; genome mining and metabolic manipulation, *Molecules* 25 (2020) 3000, <https://doi.org/10.3390/molecules25133000>.
- [12] S.A. Petropoulos, Á. Fernandes, M.I. Dias, C. Pereira, R. Calhelha, F. Di Gioia, N. Tzortzakos, M. Ivanov, M. Sokovic, L. Barros, I.C.F.R. Ferreira, Wild and cultivated centaurea raphanina subsp. Mixta: a valuable source of bioactive compounds, *Antioxidants* 9 (2020) 314, <https://doi.org/10.3390/antiox9040314>.
- [13] M. Ivanović, M.I. Razboršek, M. Kolar, Innovative extraction techniques using deep eutectic solvents and analytical methods for the isolation and characterization of natural bioactive compounds from plant material, *Plants* 9 (2020) 1–29, <https://doi.org/10.3390/plants9111428>.
- [14] C.J. Seneviratne, T. Suriyanarayanan, A.S. Widyarman, L.S. Lee, M. Lau, J. Ching, C. Delaney, G. Ramage, Multi-omics tools for studying microbial biofilms: current perspectives and future directions, *Crit. Rev. Microbiol.* 46 (2020) 759–778, <https://doi.org/10.1080/1040841X.2020.1828817>.
- [15] R. Gallego, M. Bueno, M. Herrero, Sub- and supercritical fluid extraction of bioactive compounds from plants, food-by-products, seaweeds and microalgae – an update, *TrAC Trends Anal. Chem.* 116 (2019) 198–213, <https://doi.org/10.1016/j.trac.2019.04.030>.
- [16] E. Alshafiqi, K. Begg, I. Amelio, N. Raulf, P. Lucarelli, T. Sauter, M. Tavassoli, Clinical update on head and neck cancer: molecular biology and ongoing challenges, *Cell Death Dis.* 10 (2019) 1–17, <https://doi.org/10.1038/s41419-019-1769-9>.
- [17] F.R. Pinu, D.J. Beale, A.M. Paten, K. Kouremenos, S. Swarup, H.J. Schirra, D. Wishart, Systems biology and multi-omics integration: viewpoints from the metabolomics research community, *Metabolites* 9 (2019) 76, <https://doi.org/10.3390/metabo9040076>.
- [18] Z. Peng, J.V. Bredeson, G.A. Wu, S. Shu, N. Rawat, D. Du, S. Parajuli, Q. Yu, Q. You, D.S. Rokhsar, F.G. Gmitter, Z. Deng, A chromosome-scale reference genome of trifoliolate orange (*Poncirus trifoliata*) provides insights into disease resistance, cold tolerance and genome evolution in Citrus, *Plant J.* 104 (2020) 1215–1232, <https://doi.org/10.1111/tpj.14993>.
- [19] D.B. Marchant, E.B. Sessa, P.G. Wolf, K. Heo, W.B. Barbazuk, P.S. Soltis, D. E. Soltis, The C-Fern (*Ceratopteris richardii*) genome: insights into plant genome evolution with the first partial homosporous fern genome assembly, *Sci. Rep.* 9 (2019) 1–14, <https://doi.org/10.1038/s41598-019-53968-8>.
- [20] J. Shi, L. Zhao, B. Yan, Y. Zhu, H. Ma, W. Chen, S. Ruan, Comparative transcriptome analysis reveals the transcriptional alterations in growth-and development-related genes in sweet potato plants infected and non-infected by SPFMV, SPV2, and SPVG, *IJMS* 20 (2019) 1012, <https://doi.org/10.3390/ijms20051012>.
- [21] L. Li, Y. Long, H. Li, X. Wu, Comparative transcriptome analysis reveals key pathways and hub genes in rapeseed during the early stage of *Plasmodiophora brassicae* infection, *Front. Genet.* 10 (2020) 1275, <https://doi.org/10.3389/fgene.2019.01275>.
- [22] I. Bludau, R. Aebbersold, Proteomic and interactomic insights into the molecular basis of cell functional diversity, *Nat. Rev. Mol. Cell Biol.* 21 (2020) 327–340, <https://doi.org/10.1038/s41580-020-0231-2>.
- [23] Y. Li, D. Kong, Y. Fu, M.R. Sussman, H. Wu, The effect of developmental and environmental factors on secondary metabolites in medicinal plants, *Plant Physiol. Biochem.* 148 (2020) 80–89, <https://doi.org/10.1016/j.plaphy.2020.01.006>.
- [24] M.A. Salem, L.P. De Souza, A. Serag, A.R. Fernie, M.A. Farag, S.M. Ezzat, S. Alsekh, Metabolomics in the context of plant natural products research: from sample preparation to metabolite analysis, *Metabolites* 10 (2020) 37, <https://doi.org/10.3390/metabo10010037>.
- [25] Y. Teimoori-Boghsani, A. Ganjeali, T. Cernava, H. Müller, J. Asili, G. Berg, Endophytic fungi of native salvia abrotanoides plants reveal high taxonomic diversity and unique profiles of secondary metabolites, *Front. Microbiol.* 10 (2020) 3013, <https://doi.org/10.3389/fmicb.2019.03013>.
- [26] A.H. Emwas, R. Roy, R.T. McKay, L. Tenori, E. Saccenti, G.A. Nagana Gowda, D. Raftery, F. Alahmari, L. Jaremko, M. Jaremko, D.S. Wishart, Nmr spectroscopy for metabolomics research, *Metabolites* 9 (2019) 123, <https://doi.org/10.3390/metabo9070123>.
- [27] J.T. Chen, Phytochemical omics in medicinal plants, *Biomolecules* 10 (2020) 1–7, <https://doi.org/10.3390/biom10060936>.
- [28] R. Maghembe, D. Damian, A. Makaranga, S.S. Nyandoro, S.L. Lyantagaye, S. Kusari, R. Hatti-Kaul, Omics for bioprospecting and drug discovery from bacteria and microalgae, *Antibiotics* 9 (2020) 229, <https://doi.org/10.3390/antibiotics9050229>.
- [29] B.C. Sorkin, A.J. Kuszak, G. Bloss, N.K. Fukagawa, F.A. Hoffman, M. Jafari, B. Barretti, P.N. Brown, F.D. Bushman, S.J. Casper, F.H. Chilton, C.S. Coffey, M. G. Ferruzzi, D.C. Hopp, M. Kiely, D. Lakens, J.B. MacMillan, D.O. Meltzer, M. Pator, J. Paul, K. Pritchett-Corning, S.K. Quinney, B. Rehermann, K.D. R. Sechell, N.S. Sipes, J.M. Stephens, D.L. Taylor, H. Tiriac, M.A. Walters, D. Xi, G. Zappala, G.F. Pauli, Improving natural product research translation: from source to clinical trial, *FASEB J.* 34 (2020) 41–65, <https://doi.org/10.1096/fj.201902143R>.
- [30] S. Tyanova, J. Cox, Perseus: a bioinformatics platform for integrative analysis of proteomics data in cancer research, *Methods Mol. Biol.* 1711 (2018) 133–148, https://doi.org/10.1007/978-1-4939-7493-1_7.
- [31] Z. Ye, F. Wang, F. Yan, L. Wang, B. Li, T. Liu, F. Hu, M. Jiang, W. Li, Z. Fu, Bioinformatic identification of candidate biomarkers and related transcription factors in nasopharyngeal carcinoma, *World J. Surg. Oncol.* 17 (2019) 60, <https://doi.org/10.1186/s12957-019-1605-9>.
- [32] F. Majolo, L.K. de Oliveira Becker Delwing, D.J. Marmitt, I.C. Bustamante-Filho, M.A. Goettter, Medicinal plants and bioactive natural compounds for cancer treatment: important advances for drug discovery, *Phytochem. Lett.* 31 (2019) 196–207, <https://doi.org/10.1016/j.phyto.2019.04.003>.
- [33] G. Karthikeyan, M.K. Swamy, M.R. Viknesh, R. Shurya, N. Sudhakar, Bioactive phytochemicals to fight against antimicrobial resistance. *Plant-Derived Bioact. Prod. Prop. Ther. Appl.*, Springer, Singapore, 2020, pp. 335–381, https://doi.org/10.1007/978-981-15-1761-7_14.
- [34] M.R. Howes, C.L. Quave, J. Collemare, E.C. Tatsis, D. Twilley, E. Lulekal, A. Farlow, L. Li, M. Cazar, D.J. Leaman, T.A.K. Prescott, W. Milliken, C. Martin, M.N. De Canha, N. Lall, H. Qin, B.E. Walker, C. Vásquez-Londoño, B. Allkin, M. Rivers, M.S.J. Simmonds, E. Bell, A. Battison, J. Felix, F. Forest, C. Leon, C. Williams, E. Nic Lughadha, Molecules from nature: reconciling biodiversity conservation and global healthcare imperatives for sustainable use of medicinal plants and fungi, *Plants People Planet* 2 (2020) 463–481, <https://doi.org/10.1002/ppp3.10138>.
- [35] T.A. Begenio, A.E. Tekka, T.A. Bafa, W.B. Nassir, Phytochemical investigation and characterization on the leaf extract of *Prunus africana*, *IRJPAC* (2020) 47–57, <https://doi.org/10.9734/irjpac/2020/v21i1430246>.
- [36] Q.Y. Wei, K.M. He, J.L. Chen, Y.M. Xu, A.T.Y. Lau, Phytofabrication of nanoparticles as novel drugs for anticancer applications, *Molecules* 24 (2019) 4246, <https://doi.org/10.3390/molecules24234246>.

- [37] K. Uswanthim, P. Wisitpongpan, T. Luetragoon, Molecular identification of phytochemical for anticancer treatment, *Anticancer. Agents Med. Chem.* 20 (2020) 651–666, <https://doi.org/10.2174/1871520620666200213110016>.
- [38] A. Jose, E. Kannan, P.R.A.V. Kumar, S.R.V. Madhupantula, Therapeutic potential of phytochemicals isolated from *Simarouba glauca* for inhibiting cancers: a review, *Syst. Rev. Pharm.* 10 (2019) 73–80, <https://doi.org/10.5530/srp.2019.1.12>.
- [39] J. Wang, Y.T. Liu, L. Xiao, L. Zhu, Q. Wang, T. Yan, Anti-Inflammatory effects of apigenin in lipopolysaccharide-induced inflammatory in acute lung injury by suppressing COX-2 and NF- κ B pathway, *Inflammation* 37 (2014) 2085–2090, <https://doi.org/10.1007/s10753-014-9942-x>.
- [40] Y. Yu, Y. Zhao, F. Teng, J. Li, Y. Guan, J. Xu, X. Lv, F. Guan, M. Zhang, L. Chen, Berberine improves cognitive deficiency and muscular dysfunction via activation of the AMPK/SIRT1/PGC-1 α pathway in skeletal muscle from naturally aging rats, *J. Nutr. Health Aging* 22 (2018) 710–717, <https://doi.org/10.1007/s12603-018-1015-7>.
- [41] G.E.S. Batiha, A.M. Beshbishy, L.M. Alkazmi, E.H. Nadwa, E.K. Rashwan, N. Yokoyama, I. Igarashi, In vitro and in vivo growth inhibitory activities of cryptolepine hydrate against several babesia species and theileria equi, *PLoS Negl. Trop. Dis.* 14 (2020) 1–15, <https://doi.org/10.1371/journal.pntd.0008489>.
- [42] S. Belenahalli Shekarappa, S. Kandagalla, V.H. Malojirao, P.K. Pavan, P. B.T, M. Hanumanthappa, A systems biology approach to identify the key targets of curcumin and capsaicin that downregulate pro-inflammatory pathways in human monocytes, *Comput. Biol. Chem.* 83 (2019), 107162, <https://doi.org/10.1016/j.compbiolchem.2019.107162>.
- [43] W. Zhang, W. Zhang, L. Sun, L. Xiang, X. Lai, Q. Li, S. Sun, The effects and mechanisms of epigallocatechin-3-gallate on reversing multidrug resistance in cancer, *Trends Food Sci. Technol.* 93 (2019) 221–233, <https://doi.org/10.1016/j.tifs.2019.09.017>.
- [44] D. Cox-Georgian, N. Ramadoss, C. Dona, C. Basu, Therapeutic and medicinal uses of terpenes. *Med. Plants From Farm to Pharm.*, Springer International Publishing, 2019, pp. 333–359, https://doi.org/10.1007/978-3-030-31269-5_15.
- [45] A. Fahmi, N. Hassanen, M. Abdur-Rahman, E. Shams-Eldin, Phytochemicals, antioxidant activity and hepatoprotective effect of ginger (*Zingiber officinale*) on diethylnitrosamine toxicity in rats, *Biomarkers* 24 (2019) 436–447, <https://doi.org/10.1080/1354750X.2019.1606280>.
- [46] S. So, S. Uriyapongson, J. Uriyapongson, Effects of dried tomato waste powder levels on lycopene content, lipid oxidation, color, antioxidant activity, and sensory properties of frankfurter sausage made from Thai native B Effects of dried tomato waste powder levels on lycopene content, lipid oxi, *Songklanakarin J. Sci. Technol.* 42 (2020) 27–34, <https://doi.org/10.14456/sjst-psu.2020.5>.
- [47] F. Liu, L.Y. Wang, M.C. Yu, Y.T. Li, Z.Y. Wu, C.W. Yan, A new crystalline isoniazid-quecetin with hepatoprotective effect: the design, structure, and in vitro/in vivo performance evaluation, *Eur. J. Pharm. Sci.* 144 (2020), 105216, <https://doi.org/10.1016/j.ejps.2020.105216>.
- [48] T. Farkhondeh, S.L. Folgado, A.M. Pourbagher-Shahri, M. Ashrafzadeh, S. Samarghandian, The therapeutic effect of resveratrol: focusing on the Nrf2 signaling pathway, *Biomed. Pharmacother.* 127 (2020), 110234, <https://doi.org/10.1016/j.biopha.2020.110234>.
- [49] M. Iranshahi, F. Farhadi, B. Paknejad, P. Zareian, M. Iranshahi, M. Karami, S. R. Abtahi, Gummosin, a sesquiterpene coumarin from *Ferula assa-foetida* is preferentially cytotoxic to human breast and prostate cancer cell lines, *Avicenna J. Phytomed.* 9 (2019) 446–453, <https://doi.org/10.22038/AJP.2019.12598>.
- [50] Z.Q. Zheng, W.J. Wei, J. Zhang, H.Y. Li, K. Xu, J. Xu, B. Tang, Y. Li, K. Gao, A.-E. Heliaquanoids, Five Sesquiterpenoid Dimers from *Inula helianthus-aquatica*, *J. Org. Chem.* 84 (2019) 4473–4477, <https://doi.org/10.1021/acs.joc.8b03284>.
- [51] M. Bacanli, Limonene and ursolic acid in the treatment of diabetes. *Diabetes*, Elsevier, 2020, pp. 275–283, <https://doi.org/10.1016/b978-0-12-815776-3.00027-9>.
- [52] W. Vanden Berghe, L. Sabbe, M. Kailch, G. Haegeman, K. Heynincx, Molecular insight in the multifunctional activities of Withaferin A, *Biochem. Pharmacol.* 84 (2012) 1282–1291, <https://doi.org/10.1016/j.bcp.2012.08.027>.
- [53] A. Antonini, P. Jenner, Apomorphine infusion in advanced Parkinson disease, *Nat. Rev. Neurol.* 14 (2018) 693–694, <https://doi.org/10.1038/s41582-018-0083-y>.
- [54] N.F. Kane, Effect of extracts of *Artemisia afra* collected from five different regions in Africa (Kenya, Burundi, Tanzania, South Africa and Senegal) on in vitro and in vivo cultures of *Plasmodium* Species, (2019).
- [55] G.S. Santos, S.B.P. Sinoti, F.T.C. de Almeida, D. Silveira, L.A. Simeoni, K.K. P. Gomes-Copeland, Use of galantamine in the treatment of Alzheimer's disease and strategies to optimize its biosynthesis using the in vitro culture technique, *Plant Cell. Tissue Organ Cult.* 143 (2020) 13–29, <https://doi.org/10.1007/s11240-020-01911-5>.
- [56] R. Ahmed, M. Tariq, I.S. Ahmad, H. Fouly, Fakhar-I-Abbas, A. Hasan, M. Kushad, Poly(lactic-co-glycolic acid) nanoparticles loaded with callistemon citrinus phenolics exhibited anticancer properties against three breast cancer cell lines, *J. Food Qual.* 2019 (2019) 14–15, <https://doi.org/10.1155/2019/2638481>.
- [57] T.M. Khing, W.W. Po, U.D. Sohn, Fluoxetine enhances anti-tumor activity of paclitaxel in gastric adenocarcinoma cells by triggering apoptosis and necroptosis, *Anticancer Res.* 39 (2019) 6155–6163, <https://doi.org/10.21873/anticancer.13823>.
- [58] L. Mansfield, J.A. Bernstein, Tiotropium in asthma: from bench to bedside, *Respir. Med.* 154 (2019) 47–55, <https://doi.org/10.1016/j.rmed.2019.06.008>.
- [59] F. Bray, J. Ferlay, I. Soerjomataram, R.L. Siegel, L.A. Torre, A. Jemal, Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries, *CA Cancer J. Clin.* 68 (2018) 394–424, <https://doi.org/10.3322/caac.21492>.
- [60] M.H. Saad, E.M. El-Fakharany, M.S. Salem, N.M. Sidkey, The use of cyanobacterial metabolites as natural medical and biotechnological tools: review article, *J. Biomol. Struct. Dyn.* (2020) 1–23, <https://doi.org/10.1080/07391102.2020.1838948>.
- [61] S. Gezici, N. Şekeröğlu, Current perspectives in the application of medicinal plants against cancer: novel therapeutic agents, *Anticancer Agents Med. Chem.* 19 (2019) 101–111, <https://doi.org/10.2174/1871520619666181224121004>.
- [62] A. Maruca, R. Catalano, D. Bagetta, F. Mesiti, F.A. Ambrosio, I. Romeo, F. Moraca, R. Rocca, F. Ortuso, A. Artese, G. Costa, S. Alcaro, A. Lupia, The Mediterranean Diet as source of bioactive compounds with multi-targeting anti-cancer profile, *Eur. J. Med. Chem.* 181 (2019), 111579, <https://doi.org/10.1016/j.ejmech.2019.111579>.
- [63] A. Kocyigit, E.M. Guler, M. Dikilitas, Role of antioxidant phytochemicals in prevention, formation and treatment of cancer, *React. Oxy. Species Living Cells* (2018), <https://doi.org/10.5772/intechopen.72217>.
- [64] A. Dave, F. Parande, E.-J. Park, J.M. Pezzuto, Phytochemicals and cancer chemoprevention, *J. Cancer Metastasis Treat.* 2020 (2020), <https://doi.org/10.20517/2394-4722.2020.106>.
- [65] A.T. Mbaveng, C.G.T. Noulala, A.R.M. Samba, S.B. Tankeo, G.W. Fotso, E. N. Happi, B.T. Ngadjui, V.P. Beng, V. Kuete, T. Efferth, Cytotoxicity of botanicals and isolated phytochemicals from *Araliopsis soyauxii* Engl. (Rutaceae) towards a panel of human cancer cells, *J. Ethnopharmacol.* 267 (2021), 113535, <https://doi.org/10.1016/j.jep.2020.113535>.
- [66] H. Babich, M.E. Krupka, H.A. Nissim, H.L. Zuckerbraun, Differential in vitro cytotoxicity of (-)-epicatechin gallate (ECG) to cancer and normal cells from the human oral cavity, *Toxicol. Vitro* 19 (2005) 231–242, <https://doi.org/10.1016/j.tiv.2004.09.001>.
- [67] M. Lazarević, M. Milošević, N. Petrović, S. Petrović, G. Damante, J. Milasin, B. Milovanović, Cytotoxic effects of different aromatic plants essential oils on oral squamous cell carcinoma- an in vitro study, *Balk. J. Dent. Med.* 23 (2019) 73–79, <https://doi.org/10.2478/bjdm-2019-0014>.
- [68] L. Ma, Y. Chen, R. Han, S. Wang, Benzyl isothiocyanate inhibits invasion and induces apoptosis via reducing S100A4 expression and increases PUMA expression in oral squamous cell carcinoma cells, *Braz. J. Med. Biol. Res.* 52 (2019) 8409, <https://doi.org/10.1590/1414-431x20198409>.
- [69] K. Min, T.K. Kwon, Anticancer effects and molecular mechanisms of epigallocatechin-3-gallate, *Integr. Med. Res.* 3 (2014) 16–24, <https://doi.org/10.1016/j.imr.2013.12.001>.
- [70] V. Kapoor, S. Aggarwal, S.N. Das, 6-Gingerol mediates its anti tumor activities in human oral and cervical cancer cell lines through apoptosis and cell cycle arrest, *Phytother. Res.* 30 (2016) 588–595, <https://doi.org/10.1002/ptr.5561>.
- [71] Y.T. Yeh, Y.N. Hsu, S.Y. Huang, J.S. Lin, Z.F. Chen, N.H. Chow, S.H. Su, H. W. Shyu, C.C. Lin, W.T. Huang, H. Yeh, Y. chia chih, Y.H. Huang, S.J. Su, Benzyl isothiocyanate promotes apoptosis of oral cancer cells via an acute redox stress-mediated DNA damage response, *Food Chem. Toxicol.* 97 (2016) 336–345, <https://doi.org/10.1016/j.fct.2016.09.028>.
- [72] J. Douaither, A. Ravipati, B. Grams, S. Chowdhury, O. Alatis, C. Are, Colorectal cancer—global burden, trends, and geographical variations, *J. Surg. Oncol.* 115 (2017) 619–630, <https://doi.org/10.1002/jso.24578>.
- [73] K.D. Miller, L. Nogueira, A.B. Mariotto, J.H. Rowland, K.R. Yabroff, C.M. Alfano, A. Jemal, J.L. Kramer, R.L. Siegel, Cancer treatment and survivorship statistics, 2019, *CA Cancer J. Clin.* 69 (2019) 363–385, <https://doi.org/10.3322/caac.21565>.
- [74] A.S. Choudhari, P.C. Mandave, M. Deshpande, P. Ranjekar, O. Prakash, Phytochemicals in cancer treatment: from preclinical studies to clinical practice, *Front. Pharmacol.* 10 (2020) 1614, <https://doi.org/10.3389/fphar.2019.01614>.
- [75] A. Steinbrueck, A.C. Sedgwick, J.T. Brewster, K.C. Yan, Y. Shang, D.M. Knoll, G. I. Vargas-Zúñiga, X.P. He, H. Tian, J.L. Sessler, Transition metal chelators, pro-chelators, and ionophores as small molecule cancer chemotherapeutic agents, *Chem. Soc. Rev.* 49 (2020) 3726–3747, <https://doi.org/10.1039/c9cs00373h>.
- [76] G.L. Chou, S.F. Peng, C.L. Liao, H.C. Ho, K.W. Lu, J.C. Lien, M.J. Fan, K.C. La, J. G. Chung, Casticin impairs cell growth and induces cell apoptosis via cell cycle arrest in human oral cancer SCC-4 cells, *Environ. Toxicol.* 33 (2018) 127–141, <https://doi.org/10.1002/tox.22497>.
- [77] S.M. Hu, X.H. Yao, Y.H. Hao, A.H. Pan, X.W. Zhou, 8-Gingerol regulates colorectal cancer cell proliferation and migration through the EGFR/STAT/ERK pathway, *Int. J. Oncol.* 56 (2020) 390–397, <https://doi.org/10.3892/ijo.2019.4934>.
- [78] N. Amirzargar, S. Heidari-Soureshjani, Q. Yang, S. Abbaszadeh, M. Khaksarian, Neuroprotective effects of medicinal plants in cerebral hypoxia and anoxia: a systematic review, *Nat. Prod. J.* 10 (2019) 550–565, <https://doi.org/10.2174/2210315509666190820103658>.
- [79] K.W. Luo, Wei Chen, W.Y. Lung, X.Y. Wei, B.H. Cheng, Z.M. Cai, W.R. Huang, EGCG inhibited bladder cancer SW780 cell proliferation and migration both in vitro and in vivo via down-regulation of NF- κ B and MMP-9, *J. Nutr. Biochem.* 41 (2017) 56–64, <https://doi.org/10.1016/j.jnutbio.2016.12.004>.
- [80] C.J. Cho, C.P. Yu, C.L. Wu, J.Y. Ho, C.W. Yang, D.S. Yu, Decreased drug resistance of bladder cancer using phytochemicals treatment, *Kaohsiung J. Med. Sci.* 37 (2020) 128–135, <https://doi.org/10.1002/kjms.12306>.
- [81] R. Zhang, Z. Wang, W. You, F. Zhou, Z. Guo, K. Qian, Y. Xiao, X. Wang, Suppressive effects of plumbagin on the growth of human bladder cancer cells via PI3K/AKT/mTOR signaling pathways and EMT, *Cancer Cell Int.* 20 (2020) 520, <https://doi.org/10.1186/s12935-020-01607-y>.

- [82] X. Guo, W. Zhang, J. Ren, Y. Chen, J. Wang, C. Zhu, Y. Zhang, A. Gao, LncRNA-OBFC2A targeted to Smad3 regulated Cyclin D1 influences cell cycle arrest induced by 1,4-benzoquinone, *Toxicol. Lett.* 332 (2020) 74–81, <https://doi.org/10.1016/j.toxlet.2020.07.004>.
- [83] X. Kong, J. Luo, T. Xu, Y. Zhou, Z. Pan, Y. Xie, L. Zhao, Y. Lu, X. Han, Z. Li, L. Liu, Plumbagin enhances TRAIL-induced apoptosis of human leukemic Kasumi-1 cells through upregulation of TRAIL death receptor expression, activation of caspase-8 and inhibition of cFLIP, *Oncol. Rep.* 37 (2017) 3423–3432, <https://doi.org/10.3892/or.2017.5627>.
- [84] M. Ghasemi-Pirbaluti, B. Pourghesari, H. Shirzad, Z. Sourani, P. Beshkar, The inhibitory effect of epigallocatechin gallate on the viability of T lymphoblastic leukemia cells is associated with increase of Caspase-3 level and fas expression, *Indian J. Hematol. Blood Transfus.* 34 (2018) 253–260, <https://doi.org/10.1007/s12288-017-0854-4>.
- [85] G. Zhu, Q. Shen, H. Jiang, O. Ji, L. Zhu, L. Zhang, Curcumin inhibited the growth and invasion of human monocytic leukemia SHI-1 cells in vivo by altering MAPK and MMP signalling, *Pharm. Biol.* 58 (2020) 25–34, <https://doi.org/10.1080/13880209.2019.1701042>.
- [86] H.F. Saleem, R.M. Kharshoum, H.A. Abou-Taleb, D.M. Naguib, Nanosized transferosome-based intranasal in situ gel for brain targeting of resveratrol: formulation, optimization, in vitro evaluation, and in vivo pharmacokinetic study, *AAPS PharmSciTech* 20 (2019) 181, <https://doi.org/10.1208/s12249-019-1353-8>.
- [87] A. Cano, M. Ettcheto, J.H. Chang, E. Barroso, M. Espina, B.A. Kühne, M. Barenys, C. Auladell, J. Folch, E.B. Souto, A. Camins, P. Turowski, M.L. Garcia, Dual-drug loaded nanoparticles of Epigallocatechin-3-gallate (EGCG)/Ascorbic acid enhance therapeutic efficacy of EGCG in an APPsw/PS1dE9 Alzheimer's disease mice model, *J. Control. Release* 301 (2019) 62–75, <https://doi.org/10.1016/j.jconrel.2019.03.010>.
- [88] A. Tomko, L. O'Leary, H. Trask, J.C. Achenbach, S.R. Hall, K.B. Goralski, L. D. Ellis, D.J. Dupré, Antitumor activity of abnormal cannabidiol and its analog O-1602 in taxol-resistant preclinical models of breast cancer, *Front. Pharmacol.* 10 (2019) 1124, <https://doi.org/10.3389/fphar.2019.01124>.
- [89] K. Li, W. Liu, Q. Zhao, C. Wu, C. Fan, H. Lai, S. Li, Combination of tanshinone IIA and doxorubicin possesses synergism and attenuation effects on doxorubicin in the treatment of breast cancer, *Phytother. Res.* 33 (2019) 1658–1669, <https://doi.org/10.1002/ptr.6353>.
- [90] X.J. Gou, H.H. Bai, L.W. Liu, H.Y. Chen, Q. Shi, L.S. Chang, M.M. Ding, Q. Shi, M. X. Zhou, W.L. Chen, L.M. Zhang, Asiatic acid interferes with invasion and proliferation of breast cancer cells by inhibiting WAVE3 activation through PI3K/AKT signaling pathway, *Biomed. Res. Int.* 2020 (2020), 1874387, <https://doi.org/10.1155/2020/1874387>.
- [91] J. Sheng, W. Shi, H. Guo, W. Long, Y. Wang, J. Qi, J. Liu, Y. Xu, The inhibitory effect of (–)-Epigallocatechin-3-Gallate on breast cancer progression via reducing SCUBE2 methylation and DNMT activity, *Molecules* 24 (2019) 2899, <https://doi.org/10.3390/molecules24162899>.
- [92] W.Q. Guo, Y.G. Chen, R.Z. Shi, K. He, J.F. Wang, J.H. Shao, J.B. Wan, J.L. Gao, 20 (S)-protopanaxadiol suppresses the abnormal granule-monocyte differentiation of hematopoietic stem cells in 4T1 breast cancer-bearing mouse, *Evid. Based Complement. Altern. Med.* 2020 (2020) 1–11, <https://doi.org/10.1155/2020/8747023>.
- [93] K.B. Karatay, A.Y. Kılçar, E. Derviş, F.Z.B. Müftüleri, Radioiodinated ginger compounds (6-gingerol and 6-shogaol) and incorporation assays on breast cancer cells, *Anticancer Agents Med. Chem.* 20 (2020) 1129–1139, <https://doi.org/10.2174/187152062066200128114215>.
- [94] Y. Ding, J. He, J. Huang, T. Yu, X. Shi, T. Zhang, G. Yan, S. Chen, C. Peng, Harmine induces anticancer activity in breast cancer cells via targeting TAZ, *Int. J. Oncol.* 54 (2019) 1995–2004, <https://doi.org/10.3892/ijo.2019.4777>.
- [95] J. Wang, S. Xie, J. Yang, H. Xiong, Y. Jia, Y. Zhou, Y. Chen, X. Ying, C. Chen, C. Ye, L. Wang, J. Zhou, The long noncoding RNA H19 promotes tamoxifen resistance in breast cancer via autophagy, *J. Hematol. Oncol.* 12 (2019) 81, <https://doi.org/10.1186/s13045-019-0747-0>.
- [96] M. Razaqhi, A. Ramazani, M. Khoobi, T. Mortezaazadeh, E.A. Aksoy, T. T. Küçükçiling, Highly fluorinated graphene oxide nanosheets for anticancer linoleic-curcumin conjugate delivery and T2-Weighted magnetic resonance imaging: In vitro and in vivo studies, *J. Drug Deliv. Sci. Technol.* 60 (2020), 101967, <https://doi.org/10.1016/j.jddst.2020.101967>.
- [97] M.A. Fouad, M.M. Sayed-Ahmed, E.A. Huwait, H.F. Hafez, A.M.M. Osman, Epigenetic immunomodulatory effect of eugenol and astaxanthin on doxorubicin cytotoxicity in hormonal positive breast Cancer cells, *BMC Pharmacol. Toxicol.* 22 (2021) 8, <https://doi.org/10.1186/s40360-021-00473-2>.
- [98] A.R. Pamungkas, P. Indrayudha, Aktivitas sitotoksik ekstrak etanol, fraksi etanol-air, etil asetat serta N-Heksana Buah Pare (Momordica charantia) pada Sel MCF-7 secara in-vitro, *Pharm. J. Farm. Indones.* 16 (2019) 73–82, <https://doi.org/10.23917/pharmacon.v16i2.9049>.
- [99] Q. Zhang, D. Yang, Allicin suppresses the migration and invasion in cervical cancer cells mainly by inhibiting NRF2, *Exp. Ther. Med.* 17 (2018) 1523–1528, <https://doi.org/10.3892/etm.2018.7104>.
- [100] S.D. Mane, A.N. Kamatham, Ascorbyl stearate stimulates cell death by oxidative stress-mediated apoptosis and autophagy in HeLa cervical cancer cell line in vitro, *3 Biotech* 9 (2019) 1–15, <https://doi.org/10.1007/s13205-019-1628-5>.
- [101] J. Stefanowicz-Hajduk, M. Gucwa, A. Hajduk, Jr. Ochocka, Kalanchoe blossfeldiana extract induces cell cycle arrest and necrosis in human cervical cancer cells, *Pharmacogn. Mag.* 15 (2019) 527, https://doi.org/10.4103/pm_pm_86_19.
- [102] S. Mavrikou, V. Tsekouras, M.A. Karageorgou, G. Moschopoulou, S. Kintzios, Detection of superoxide alterations induced by 5-fluorouracil on HeLa cells with a cell-based biosensor, *Biosensors* 9 (2019) 126, <https://doi.org/10.3390/bios9040126>.
- [103] N. Rastogi, S. Duggal, S.K. Singh, K. Porwal, V.K. Srivastava, R. Maurya, M.L. B. Bhatt, D.P. Mishra, Proteasome inhibition mediates p53 reactivation and anticancer activity of 6-Gingerol in cervical cancer cells, *Oncotarget* 6 (2015) 43310–43325, <https://doi.org/10.18632/oncotarget.6383>.
- [104] Z. Wang, M.Y. Li, Z.H. Zhang, H.X. Zuo, J.Y. Wang, Y. Xing, M.H. Ri, H.L. Jin, C. H. Jin, G.H. Xu, L.X. Piao, C.G. Jiang, J. Ma, X. Jin, Panaxadiol inhibits programmed cell death-ligand 1 expression and tumour proliferation via hypoxia-inducible factor (HIF)-1 α and STAT3 in human colon cancer cells, *Pharmacol. Res.* 155 (2020), 104727, <https://doi.org/10.1016/j.phrs.2020.104727>.
- [105] H. Zhang, N. Cao, Z. Yang, X. Fang, X. Yang, H. Li, Z. Hong, Z. Ji, Bilobalide alleviated dextran sulfate sodium-induced experimental colitis by inhibiting M1 macrophage polarization through the NF- κ B signaling pathway, *Front. Pharmacol.* 11 (2020) 718, <https://doi.org/10.3389/fphar.2020.00718>.
- [106] L. Wang, W. Huang, J. Zhan, Grape seed proanthocyanidins induce autophagy and modulate survivin in HepG2 cells and inhibit xenograft tumor growth in vivo, *Nutrients* 11 (2019) 2983, <https://doi.org/10.3390/nu1122983>.
- [107] H. Yang, E. Sukamtoh, Z. Du, W. Wang, M. Ando, Y.N. Kwakwa, J. Zhang, G. Zhang, Click chemistry approach to characterize curcumin-protein interactions in vitro and in vivo, *J. Nutr. Biochem.* 68 (2019) 1–6, <https://doi.org/10.1016/j.jnutbio.2019.02.010>.
- [108] M.L. Cuellar-Núñez, G. Loarca-Piña, M. Berhow, E. Gonzalez de Mejia, Glucosinolate-rich hydrolyzed extract from Moringa oleifera leaves decreased the production of TNF- α and IL-1 β cytokines and induced ROS and apoptosis in human colon cancer cells, *J. Funct. Foods* 75 (2020), 104270, <https://doi.org/10.1016/j.jff.2020.104270>.
- [109] Y.M. Lin, C.I. Chen, Y.P. Hsiang, Y.C. Hsu, K.C. Cheng, P.H. Chien, H.L. Pan, C. C. Lu, Y.J. Chen, Chrysin attenuates cell viability of human colorectal cancer cells through autophagy induction unlike 5-fluorouracil/oxaliplatin, *Int. J. Mol. Sci.* 19 (2018) 1763, <https://doi.org/10.3390/ijms19061763>.
- [110] Y. Yao, C. Rao, G. Zheng, S. Wang, Luteolin suppresses colorectal cancer cell metastasis via regulation of the miR-384/pleiotrophin axis, *Oncol. Rep.* 42 (2019) 131–141, <https://doi.org/10.3892/or.2019.7136>.
- [111] Z. Wu, X. Han, G. Tan, Q. Zhu, H. Chen, Y. Xia, J. Gong, Z. Wang, Y. Wang, J. Yan, Dioscin inhibited glycolysis and induced cell apoptosis in colorectal cancer via promoting c-myc ubiquitination and subsequent hexokinase-2 suppression, *Oncotargets Ther.* 13 (2020) 31–44, <https://doi.org/10.2147/OTT.S224062>.
- [112] C.S. Lei, Y.C. Hou, M.H. Pai, M.T. Lin, S.L. Yeh, Effects of quercetin combined with anticancer drugs on metastasis-associated factors of gastric cancer cells: in vitro and in vivo studies, *J. Nutr. Biochem.* 51 (2018) 105–113, <https://doi.org/10.1016/j.jnutbio.2017.09.011>.
- [113] C. Eder-Czembirek, B.M. Erovic, C. Czembirek, M. Brunner, E. Selzer, R. Pötter, D. Thurnher, Betulinic acid a radiosensitizer in head and neck squamous cell carcinoma cell lines, *Strahlenther. Onkol.* 186 (2010) 143–148, <https://doi.org/10.1007/s00066-010-2069-6>.
- [114] K.C. Lai, F.S. Chueh, Y.T. Hsiao, Z.Y. Cheng, J.C. Lien, K.C. Liu, S.F. Peng, J. G. Chung, Gefitinib and curcumin-loaded nanoparticles enhance cell apoptosis in human oral cancer SAS cells in vitro and inhibit SAS cell xenografted tumor in vivo, *Toxicol. Appl. Pharmacol.* 382 (2019), 114734, <https://doi.org/10.1016/j.taap.2019.114734>.
- [115] C. Karavasili, D.A. Andreadis, O.L. Katsamenis, E. Panteris, P. Anastasiadou, Z. Kakazanis, V. Zoumpourlis, C.K. Markopoulou, S. Koutsopoulos, I. S. Vizirianakis, D.G. Fatouros, Synergistic antitumor potency of a self-assembling peptide hydrogel for the local co-delivery of doxorubicin and curcumin in the treatment of head and neck cancer, *Mol. Pharm.* 16 (2019) 2326–2341, <https://doi.org/10.1021/acs.molpharmaceut.8b01221>.
- [116] G.T. Bolger, A. Licollari, R. Bagshaw, A. Tan, R. Greil, B. Vcelar, M. Majeed, P. Sordillo, Intense uptake of liposomal curcumin by multiple myeloma cell lines: Comparison to normal lymphocytes, red blood cells and chronic lymphocytic leukemia cells, *Anticancer Res.* 39 (2019) 1161–1168, <https://doi.org/10.21873/anticancer.13225>.
- [117] Y. yang Ma, Z. min Di, Q. Cao, W. shuang Xu, S. xing Bi, J. shuang Yu, Y. jun Shen, Y. qiang Yu, Y. xian Shen, L. jie Feng, Xanthin induces glioma cell apoptosis and inhibits tumor growth via activating endoplasmic reticulum stress-dependent CHOP pathway, *Acta Pharmacol. Sin.* 41 (2020) 404–414, <https://doi.org/10.1038/s41401-019-0318-5>.
- [118] S.S. Park, H. Ohba, Suppressive activity of protease inhibitors from buckwheat seeds againsts human t-acute lymphoblastic leukemia cell lines, *Appl. Biochem. Biotechnol. Part A Enzym. Eng. Biotechnol.* 117 (2004) 65–74, <https://doi.org/10.1385/abab:117:2:065>.
- [119] G.J. Huang, M.J. Sheu, H.J. Chen, Y.S. Chang, Y.H. Lin, Growth inhibition and induction of apoptosis in NB4 promyelocytic leukemia cells by trypsin inhibitor from sweet potato storage roots, *J. Agric. Food Chem.* 55 (2007) 2548–2553, <https://doi.org/10.1021/jf063008m>.
- [120] E. Nibret, M. Youns, R.L. Krauth-Siegel, M. Wink, Biological activities of xanthin from Xanthium strumarium leaves, *Phytother. Res.* 25 (2011) 1883–1890, <https://doi.org/10.1002/ptr.3651>.
- [121] Z. Mao, X. Han, D. Chen, Y. Xu, L. Xu, L. Yin, H. Sun, Y. Qi, L. Fang, K. Liu, J. Peng, Potent effects of dioscin against hepatocellular carcinoma through regulating TP53-induced glycolysis and apoptosis regulator (TIGAR)-mediated apoptosis, autophagy, and DNA damage, *Br. J. Pharmacol.* 176 (2019) 919–937, <https://doi.org/10.1111/bph.14594>.

- [122] J. Li, H. Li, Y. Yu, Y. Liu, Y. Liu, Q. Ma, L. Zhang, X. Lu, X. yang Wang, Z. Chen, D. Zuo, J. Zhou, Mannan-binding lectin suppresses growth of hepatocellular carcinoma by regulating hepatic stellate cell activation via the ERK/COX-2/PGE 2 pathway, *Oncoimmunology* 8 (2019), 1527650, <https://doi.org/10.1080/2162402X.2018.1527650>.
- [123] D. Zhang, J. Zhang, J. Zeng, Z. Li, H. Zuo, C. Huang, X. Zhao, Nano-gold loaded with resveratrol enhance the anti-hepatoma effect of resveratrol in vitro and in vivo, *J. Biomed. Nanotechnol.* 15 (2019) 288–300, <https://doi.org/10.1166/jbn.2019.2682>.
- [124] M. Fathy, M. Okabe, E.M. Othman, H.M. Saad Eldien, T. Yoshida, Preconditioning of adipose-derived mesenchymal stem-like cells with eugenol potentiates their migration and proliferation in vitro and therapeutic abilities in rat hepatic fibrosis, *Molecules* 25 (2020) 2020, <https://doi.org/10.3390/molecules25092020>.
- [125] M.Q. Pham, T.H. Van Tran, Q.L. Pham, J.E. Gairin, In silico analysis of the binding properties of solasonine to mortalalin and p53, and in vitro pharmacological studies of its apoptotic and cytotoxic effects on human HepG2 and Hep3b hepatocellular carcinoma cells, *Fundam. Clin. Pharmacol.* 33 (2019) 385–396, <https://doi.org/10.1111/fcp.12447>.
- [126] Y. Lei, H. Gan, Y. Huang, Y. Chen, L. Chen, A. Shan, H. Zhao, M. Wu, X. Li, Q. Ma, J. Wang, E. Zhang, J. Zhang, Y. Li, F. Xue, L. Deng, Digitoxin inhibits proliferation of multidrug-resistant HepG2 cells through G2/M cell cycle arrest and apoptosis, *Oncol. Lett.* 20 (2020) 71, <https://doi.org/10.3892/ol.2020.11932>.
- [127] G. Deng, S. Zeng, J. Ma, Y. Zhang, Y. Qu, Y. Han, L. Yin, C. Cai, C. Guo, H. Shen, The anti-tumor activities of Neferine on cell invasion and oxaliplatin sensitivity regulated by EMT via Snail signaling in hepatocellular carcinoma, *Sci. Rep.* 7 (2017) 1–14, <https://doi.org/10.1038/srep41616>.
- [128] Y.J. Chen, Z.W. Wang, T.L. Lu, C.B. Gomez, H.W. Fang, Y. Wei, C.L. Tseng, The synergistic anticancer effect of dual drug- (Cisplatin/Epigallocatechin Gallate) loaded gelatin nanoparticles for lung cancer treatment, *J. Nanomater.* 2020 (2020) 1–15, <https://doi.org/10.1155/2020/9181549>.
- [129] Y.P. Huang, Y.W. Jiang, H.Y. Chen, Y.T. Hsiao, S.F. Peng, Y.C. Chou, J.L. Yang, T. C. Hsia, J.G. Chung, Benzyl isothiocyanate induces apoptotic cell death through mitochondria-dependent pathway in gefitinib-resistant NCI-H460 human lung cancer cells in vitro, *Anticancer Res.* 38 (2018) 5165–5176, <https://doi.org/10.21873/anticancer.12839>.
- [130] A.A. Abd-Rabou, H.H. Ahmed, Bevacizumab and CCR2 inhibitor nanoparticles induce cytotoxicity-mediated apoptosis in doxorubicin-treated hepatic and non-small lung cancer cells, *Asian Pac. J. Cancer Prev.* 20 (2019) 2225–2238, <https://doi.org/10.31557/APJCP.2019.20.7.2225>.
- [131] H. Bakshi, L. Hakkim, H.A. Bakshi, S. Sam, M. Al-Buloshi, F. Lukmanul Hakkim, Assessment of in vitro cytotoxicity of saffron (*Crocus sativus* L.) on cervical cancer cells (HEp-2) and their in vivo pre-clinical toxicity in normal swiss albino mice Cancer drug discovery and cancer cell signaling View project Assessment of in vitro cyt, *Int. J. Herb. Med.* 4 (2016) 80–83.
- [132] F. Shi, C. Wang, L. Wang, X. Song, H. Yang, Q. Fu, W. Zhao, Preparative isolation and purification of steroidal glycoalkaloid from the ripe berries of *Solanum nigrum* L. by preparative HPLC-MS and UHPLC-TOF-MS/MS and its anti-non-small cell lung tumors effects in vitro and in vivo, *J. Sep. Sci.* 42 (2019) 2471–2481, <https://doi.org/10.1002/jssc.201801165>.
- [133] L. Zhou, F. Wu, W. Jin, B. Yan, X. Chen, Y. He, W. Yang, W. Du, Q. Zhang, Y. Guo, Q. Yuan, X. Dong, W. Yu, J. Zhang, L. Xiao, P. Tong, L. Shan, T. Efferth, Theabrownin inhibits cell cycle progression and tumor growth of lung carcinoma through c-myc-related mechanism, *Front. Pharmacol.* 8 (2017) 75, <https://doi.org/10.3389/fphar.2017.00075>.
- [134] V. Magesh, K.D. Bhavani, P. Senthilnathan, P. Rajendran, D. Sakthisekaran, In vivo protective effect of crocetin on benzo(a)pyrene-induced lung cancer in swiss albino mice, *Phytother. Res.* 23 (2009) 533–539, <https://doi.org/10.1002/ptr.2666>.
- [135] H.N. Oh, A.W. Kwak, M.H. Lee, E. Kim, G. Yoon, S.S. Cho, K. Liu, J. Il Chae, J. H. Shim, Targeted inhibition of c-MET by podophyllotoxin promotes caspase-dependent apoptosis and suppresses cell growth in gefitinib-resistant non-small cell lung cancer cells, *Phytomedicine* 80 (2021), 153355, <https://doi.org/10.1016/j.phymed.2020.153355>.
- [136] A. Bhaumik, S. Prasad, Studies on the antitumor potentials of betulinic acid against murine ascites Dalton's lymphoma, *Int. J. Basic Clin. Pharmacol.* (2016) 1664–1671, <https://doi.org/10.18203/2319-2003.ijbcp20162538>.
- [137] A.S.W. Ang, R.C.F. Cheung, X. Dan, Y.S. Chan, W. Pan, T.B. Ng, Purification and characterization of a glucosamine-binding antifungal lectin from *Phaseolus vulgaris* cv. Chinese Pinto Beans with antiproliferative activity towards nasopharyngeal carcinoma cells, *Appl. Biochem. Biotechnol.* 172 (2014) 672–686, <https://doi.org/10.1007/s12010-013-0542-2>.
- [138] D. Zhu, M. Shao, J. Yang, M. Fang, S. Liu, D. Lou, R. Gao, Y. Liu, A. Li, Y. Lv, Z. Mo, Q. Fan, Curcumin enhances radiosensitization of nasopharyngeal carcinoma via mediating regulation of tumor stem-like cells by a CircRNA network, *J. Cancer* 11 (2020) 2360–2370, <https://doi.org/10.7150/jca.39511>.
- [139] C.H. Holschneider, M.T. Johnson, R.M. Knox, A. Rezai, W.J. Ryan, F.J. Montz, Bullatacin - in vivo and in vitro experience in an ovarian cancer model, *Cancer Chemother. Pharmacol.* 34 (1994) 166–170, <https://doi.org/10.1007/BF00685935>.
- [140] R. Pashaei-Asl, F. Pashaei-Asl, P.M. Gharabaghi, K. Khodadadi, M. Ebrahimi, E. Ebrahimi, M. Pashaiasl, The inhibitory effect of ginger extract on Ovarian cancer cell line; application of systems biology, *Adv. Pharm. Bull.* 7 (2017) 241–249, <https://doi.org/10.15171/apb.2017.029>.
- [141] L. Xu, J. Yu, D. Zhai, D. Zhang, W. Shen, L. Bai, Z. Cai, C. Yu, Role of JNK activation and mitochondrial Bax translocation in allixin-induced apoptosis in human ovarian cancer SKOV3 cells, *Evid. Based Complement. Altern. Med.* 2014 (2014) 1–6, <https://doi.org/10.1155/2014/378684>.
- [142] C.J. Wang, C. Wang, J. Han, Y.K. Wang, L. Tang, D.W. Shen, Y. Zhao, R.H. Xu, H. Zhang, Effect of combined treatment with recombinant interleukin-2 and allixin on pancreatic cancer, *Mol. Biol. Rep.* 40 (2013) 6579–6585, <https://doi.org/10.1007/s11033-013-2766-1>.
- [143] S.O. Kim, M.R. Kim, [6]-Gingerol prevents disassembly of cell junctions and activities of MMPs in invasive human pancreas cancer cells through ERK/NF-κB/Snail signal transduction pathway, *Evid. Based Complement Altern. Med.* 2013 (2013), 761852, <https://doi.org/10.1155/2013/761852>.
- [144] C. Kürbitz, D. Heise, T. Redmer, F. Goumas, A. Arlt, J. Lemke, G. Rimbach, H. Kalthoff, A. Trauzold, Epicatechin gallate and catechin gallate are superior to epigallocatechin gallate in growth suppression and anti-inflammatory activities in pancreatic tumor cells, *Cancer Sci.* 102 (2011) 728–734, <https://doi.org/10.1111/j.1349-7006.2011.01870.x>.
- [145] L. De Petrocellis, A. Ligresti, A. Schiano Moriello, M. Iappelli, R. Verde, C.G. Stott, L. Cristino, P. Orlando, V. Di Marzo, Non-THC cannabinoids inhibit prostate carcinoma growth in vitro and in vivo: Pro-apoptotic effects and underlying mechanisms, *Br. J. Pharmacol.* 168 (2013) 79–102, <https://doi.org/10.1111/j.1476-5381.2012.02027.x>.
- [146] V. Sanna, C.K. Singh, R. Jashari, V.M. Adhami, J.C. Chamcheu, I. Rady, M. Sechi, H. Mukhtar, I.A. Siddiqui, Targeted nanoparticles encapsulating (-)-epigallocatechin-3-gallate for prostate cancer prevention and therapy, *Sci. Rep.* 7 (2017) 1–15, <https://doi.org/10.1038/srep41573>.
- [147] H.L. Tan, J.M. Thomas-Ahner, N.E. Moran, J.L. Cooperstone, J.W. Erdman, G. S. Young, S.K. Clinton, B-Carotene 90,100 oxygenase modulates the anticancer activity of dietary tomato or lycopene on prostate carcinogenesis in the TRAMP model, *Cancer Prev. Res.* 10 (2017) 161–169, <https://doi.org/10.1158/1940-6207.CCR-15-0402>.
- [148] P.J. Magee, R. Owusu-Apenten, M.J. McCann, C.I. Gill, I.R. Rowland, Chickpea (*Cicer arietinum*) and other plant-derived protease inhibitor concentrates inhibit breast and prostate cancer cell proliferation in vitro, *Nutr. Cancer* 64 (2012) 741–748, <https://doi.org/10.1080/01635581.2012.688914>.
- [149] A. Seeni, S. Takahashi, K. Takeshita, M. Tang, S. Sugiura, S.Y. Sato, T. Shirai, Suppression of prostate cancer growth by resveratrol in the transgenic for adenocarcinoma of prostate (TRAP) model, (2008).
- [150] E.B. Byun, H.M. Kim, N.Y. Sung, M.S. Yang, W.S. Kim, D.S. Choi, S. Mushtaq, S. S. Lee, E.H. Byun, Gamma irradiation of aloe-emodin induced structural modification and apoptosis through a ROS- and caspase-dependent mitochondrial pathway in stomach tumor cells, *Int. J. Radiat. Biol.* 94 (2018) 403–416, <https://doi.org/10.1080/09553002.2018.1440330>.
- [151] M. Imran, M.K. Iqbal, K. Imtiyaz, S. Saleem, S. Mittal, M.M.A. Rizvi, J. Ali, S. Baboota, Topical nanostructured lipid carrier gel of quercetin and resveratrol: formulation, optimization, in vitro and ex vivo study for the treatment of skin cancer, *Int. J. Pharm.* 587 (2020), 119705, <https://doi.org/10.1016/j.ijpharm.2020.119705>.
- [152] H.C. Pal, S.K. Katiyar, Cryptolepine, a plant alkaloid, inhibits the growth of non-melanoma skin cancer cells through inhibition of topoisomerase and induction of DNA damage, *Molecules* 21 (2016) 1758, <https://doi.org/10.3390/molecules21121758>.
- [153] B.E. Cham, Solasodine rhamnosyl glycosides specifically bind cancer cell receptors and induce apoptosis and necrosis. Treatment for skin cancer and hope for internal cancers, *Res. J. Biol. Sci.* 4 (2007) 503–514.
- [154] R. Gowda, G. Kardos, A. Sharma, S. Singh, G.P. Robertson, Nanoparticle-based celecoxib and plumbagin for the synergistic treatment of melanoma, *Mol. Cancer Ther.* 16 (2017) 440–452, <https://doi.org/10.1158/1535-7163.MCT-16-0285>.
- [155] D.A. Abdel Fadel, R. Kamel, M. Fadel, PEGylated lipid nanocarrier for enhancing photodynamic therapy of skin carcinoma using curcumin: in-vitro/in-vivo studies and histopathological examination, *Sci. Rep.* 10 (2020) 10435, <https://doi.org/10.1038/s41598-020-67349-z>.